Package 'gcrma'

July 4, 2025
Title Background Adjustment Using Sequence Information
Version 2.81.0
Author Jean(ZHIJIN) Wu, Rafael Irizarry with contributions from James MacDonald < jmacdon@med.umich.edu> Jeff Gentry
Description Background adjustment using sequence information
Maintainer Z. Wu <zwu@stat.brown.edu></zwu@stat.brown.edu>
License LGPL
Depends R (>= 2.6.0), affy (>= 1.23.2), graphics, methods, stats, utils Imports Biobase, affy (>= 1.23.2), affyio (>= 1.13.3), XVector, Biostrings (>= 2.11.32), splines, BiocManager
Suggests affydata, tools, splines, hgu95av2cdf, hgu95av2probe
biocViews Microarray, OneChannel, Preprocessing
git_url https://git.bioconductor.org/packages/gcrma
git_branch devel
git_last_commit e116657
git_last_commit_date 2025-04-15
Repository Bioconductor 3.22
Date/Publication 2025-07-04
Contents
Contents
affinity.spline.coefs bg.adjust.affinities bg.adjust.gcrma bg.parameters.ns compute.affinities fast.bkg gcrma gcrma.engine gcrma.engine2 getCDF justGCRMA

2 bg.adjust.affinities

Index 15

affinity.spline.coefs Spline coefficients for estimation of affinity from probe sequence

Description

Spline coefficients for estimation of affinity from probe sequence

Usage

```
data(affinity.spline.coefs)
```

See Also

compute.affinities

bg.adjust.affinities Background adjustment with sequence information (internal function)

Description

An internal function to be used by gcrma.

Usage

```
bg.adjust.fullmodel(pms,mms,ncs=NULL,apm,amm,anc=NULL,index.affinities,k=6
* fast + 0.25 * (1 - fast),rho=.7,fast=FALSE)
bg.adjust.affinities(pms,ncs,apm,anc,index.affinities,k=6
* fast + 0.25 * (1 - fast),fast=FALSE,nomm=FALSE)
```

Arguments

	pms	PM intensities after optical background correction, before non-specific-binding correction.
	mms	MM intensities after optical background correction, before non-specific-binding correction.
	ncs	Negative control probe intensities after optical background correction, before non-specific-binding correction. If ncs=NULL, the MM probes are considered the negative control probes.
index.affinities		
		The index of pms with known sequences. (For some types of arrays the sequences of a small subset of probes are not provided by Affymetrix.)
	apm	Probe affinities for PM probes with known sequences.
	amm	Probe affinities for MM probes with known sequences.
	anc	Probe affinities for Negative control probes with known sequences. This is ignored when ncs=NULL.
	rho	correlation coefficient of log background intensity in a pair of pm/mm probes. Default=.7

bg.adjust.gcrma 3

k A tuning parameter. See details.

fast Logical value. If TRUE a faster add-hoc algorithm is used.

nomm Logical value indicating if MM intensities are available and will to be used to

estimate background.

Details

Assumes PM=background1+signal,mm=background2, (log(background1),log(background2))' follow bivariate normal distribution, signal distribution follows power law. bg.parameters.gcrma and sg.parameters.gcrma provide adhoc estimates of the parameters.

the original gcrma uses an empirical Bayes estimate. this requires a complicated numerical integration. An add-hoc method tries to imitate the empirical Bayes estimate with a PM-B but values of PM-B<k going to k. This can be thought as a shrunken MVUE. For more details see Wu et al. (2003).

Value

a vector of same length as x.

Author(s)

Rafeal Irizarry, Zhijin(Jean) Wu

See Also

gcrma

bg.adjust.gcrma

GCRMA background adjust (internal function)

Description

This function performs background adjustment (optical noise and non-specific binding on an AffyBatch project and returns an AffyBatch object in which the PM intensities are adjusted.

```
bg.adjust.gcrma(object,affinity.info=NULL,
    affinity.source=c("reference","local"),
    NCprobe=NULL,
    type=c("fullmodel","affinities","mm","constant"),
    k=6*fast+0.5*(1-fast),stretch=1.15*fast+1*(1-fast),correction=1,
    GSB.adjust=TRUE,
    rho=.7,optical.correct=TRUE,verbose=TRUE,fast=TRUE)
```

4 bg.adjust.gcrma

Arguments

object an AffyBatch

affinity.info NULL or an AffyBatch containing the affinities in the exprs slot. This object

can be created using the function compute.affinities.

affinity.source

reference: use the package internal Non-specific binding data or local: use the experimental data in object. If local is chosen, either MM probes or a

user-defined list of probes (see NCprobes) are used to estimate affinities.

NCprobe Index of negative control probes. When set as NULL, the MM probes will be

> used. These probes are used to estimate parameters of non-specific binding on each array. These will be also used to estimate probe affinity profiles when

affinity.info is not provided.

"fullmodel" for sequence and MM model. "affinities" for sequence information type

only. "mm" for using MM without sequence information.

k A tuning factor.

stretch correction

GSB.adjust Logical value. If TRUE, probe effects in specific binding will be adjusted.

rho correlation coefficient of log background intensity in a pair of pm/mm probes.

Default=.7

optical.correct

Logical value. If TRUE, optical background correction is performed.

verbose Logical value. If TRUE messages about the progress of the function is printed.

fast Logical value. If TRUE a faster ad hoc algorithm is used.

Details

The returned value is an AffyBatch object, in which the PM probe intensities have been background adjusted. The rest is left the same as the starting AffyBatch object.

The tunning factor k will have different meanings if one uses the fast (ad hoc) algorithm or the empirical bayes approach. See Wu et al. (2003)

Value

An AffyBatch.

Author(s)

Rafeal Irizarry

Examples

```
if(require(affydata) & require(hgu95av2probe) & require(hgu95av2cdf)){
         data(Dilution)
         ai <- compute.affinities(cdfName(Dilution))</pre>
         Dil.adj<-bg.adjust.gcrma(Dilution, affinity.info=ai, type="affinities")
    }
```

bg.parameters.ns 5

bg.parameters.ns	Estimation of non-specific Binding Background Parameters
• .	v i v

Description

An internal function to be used by gcrma

Usage

bg.parameters.ns(x,affinities,affinities2=NULL,affinities3=NULL,span=.2)

Arguments

X	PM or MM intensities after optical background correction, before non-specific-binding correction.
affinities	Probe affinities for probes with known sequences. Used to estimate the function between non-specific binding and affinities.
affinities2	Probe affinities for the probes whoes expected non-specific binding intensity is to be predicted.
affinities3	Probe affinities for another extra group of probes whoes expected non-specific binding intensity is to be predicted.
span	The span parameter passed to loess function

Value

a vector of same length as x.

Author(s)

Rafeal Irizarry, Zhijin (Jean) Wu

See Also

gcrma

 $compute.affinities \qquad \textit{Probe Affinity computation}$

Description

An internal function to calculate probe affinities from their sequences.

```
compute.affinities(cdfname,verbose=TRUE)
compute.affinities2(cdfname,verbose=TRUE)
check.probes(probepackage,cdfname)
```

6 fast.bkg

Arguments

cdfname Object of class character representing the name of CDF file associated with the

arrays in the AffyBatch.

probepackage character representing the name of the package with the probe sequence infor-

mation.

verbose Logical value. If TRUE messages about the progress of the function is printed.

Details

The affinity of a probe is described as the sum of position-dependent base affinities. Each base at each position contributes to the total affinity of a probe in an additive fashion. For a given type of base, the positional effect is modeled as a spline function with 5 degrees of freedom.

Use compute.affinities2 if there are no MM probes.

check.probes makes sure things are matching as they should.

Value

compute.affinities returns an AffyBatch with the affinities for PM probes in the pm locations and the affinities for MM probes in the mm locations. NA will be added for probes with no sequence information.

Author(s)

Rafeal Irizarry

References

Hekstra, D., Taussig, A. R., Magnasco, M., and Naef, F. (2003) Absolute mRNA concentrations from sequence-specific calibration of oligonucleotide array. Nucleic Acids Research, 31. 1962-1968.

See Also

gcrma,affinity.spline.coefs

fast.bkg Internal functions for justGCRMA

Description

These are internal functions for justGCRMA that are called based on memory or speed constraints.

```
fast.bkg(filenames, pm.affinities, mm.affinities, index.affinities,
type, minimum, optical.correct, verbose, k, rho, correction, stretch,
fast, cdfname, read.verbose)
mem.bkg(filenames, pm.affinities, mm.affinities, index.affinities, type,
minimum, optical.correct, verbose, k, rho, correction, stretch, fast,
cdfname, read.verbose)
```

fast.bkg 7

Arguments

filenames A list of cel files.

pm.affinities Values passed from compute.affinities.mm.affinities Values passed from compute.affinities.

index.affinities

Values passed from compute.affinities.

type "fullmodel" for sequence and MM model. "affinities" for sequence information

only. "mm" for using MM without sequence information.

minimum A minimum value to be used for optical.correct.

optical.correct

Logical value. If TRUE, optical background correction is performed.

verbose Logical value. If TRUE, messages about the progress of the function are printed.

k A tuning factor.

rho correlation coefficient of log background intensity in a pair of pm/mm probes.

Default=.7

correction

stretch

fast Logical value. If TRUE, then a faster ad hoc algorithm is used.

cdfname Used to specify the name of an alternative cdf package. If set to NULL, the

usual cdf package based on Affymetrix' mappings will be used. Note that the name should not include the 'cdf' on the end, and that the corresponding probe package is also required to be installed. If either package is missing an error will

result.

read.verbose Logical value. If TRUE, a message is returned as each celfile is read in.

Details

Note that this expression measure is given to you in log base 2 scale. This differs from most of the other expression measure methods.

The tuning factor 'k' will have different meanings if one uses the fast (add-hoc) algorithm or the empirical Bayes approach. See Wu et al. (2003)

Value

An ExpressionSet.

Author(s)

James W. MacDonald <jmacdon@med.umich.edu>

See Also

gcrma

8 gcrma

gcrma

Robust Multi-Array expression measure using sequence information

Description

This function converts an AffyBatch into an ExpressionSet using the robust multi-array average (RMA) expression measure with help of probe sequence.

Usage

Arguments

object an AffyBatch

affinity.info NULL or an AffyBatch containing the affinities in the exprs slot. This object

can be created using the function compute.affinities.

affinity.source

reference: use the package internal Non-specific binding data or local: use the experimental data in object. If local is chosen, either MM probes or a

user-defined list of probes (see NCprobes) are used to estimate affinities.

NCprobe Index of negative control probes. When set as NULL, the MM probes will be

used. These probes are used to estimate parameters of non-specific binding on each array. These will be also used to estimate probe affinity profiles when

affinity.info is not provided.

type "fullmodel" for sequence and MM model. "affinities" for sequence information

only. "mm" for using MM without sequence information.

k A tuning factor.

stretch . correction .

GSB. adjust Logical value. If TRUE, probe effects in specific binding will be adjusted.

rho correlation coefficient of log background intensity in a pair of pm/mm probes.

Default=.7

optical.correct

Logical value. If TRUE, optical background correction is performed.

verbose Logical value. If TRUE messages about the progress of the function is printed.

fast Logical value. If TRUE a faster ad hoc algorithm is used.

subset a character vector with the the names of the probesets to be used in expression

calculation.

normalize logical value. If 'TRUE' normalize data using quantile normalization.

... further arguments to be passed (not currently implemented - stub for future use).

gcrma.engine 9

Details

Note that this expression measure is given to you in log base 2 scale. This differs from most of the other expression measure methods.

The tuning factor k will have different meanings if one uses the fast (add-hoc) algorithm or the empirical Bayes approach. See Wu et al. (2003)

Value

An ExpressionSet.

Author(s)

Rafeal Irizarry

Examples

```
if(require(affydata) & require(hgu95av2probe) & require(hgu95av2cdf)){
    data(Dilution)
    ai <- compute.affinities(cdfName(Dilution))
    Dil.expr<-gcrma(Dilution,affinity.info=ai,type="affinities")
}</pre>
```

gcrma.engine

GCRMA background adjust engine(internal function)

Description

This function adjust for non-specific binding when all arrays in the dataset share the same probe affinity information. It takes matrices of PM probe intensities, MM probe intensities, other negative control probe intensities(optional) and the associated probe affinities, and return one matrix of non-specific binding corrected PM probe intensities.

Usage

Arguments

pms	The matrix of PM intensities	
mms	The matrix of MM intensities	
ncs	The matrix of negative control probe intensities. When left asNULL, the MMs are considered the negative control probes.	
pm.affinities	The vector of PM probe affinities. Note: This can be shorter than the number of rows in pms when some probes do not have sequence information provided.	
mm.affinities	The vector of MM probe affinities.	

10 gcrma.engine2

anc The vector of Negative Control probe affinities. This is ignored if MMs are used

as negative controls (ncs=NULL)

type "fullmodel" for sequence and MM model. "affinities" for sequence information

only. "mm" for using MM without sequence information.

k A tuning factor.

stretch . correction .

GSB. adjust Logical value. If TRUE, probe effects in specific binding will be adjusted.

rho correlation coefficient of log background intensity in a pair of pm/mm probes.

Default=.7

verbose Logical value. If TRUE messages about the progress of the function is printed.

fast Logical value. If TRUE a faster add-hoc algorithm is used.

Details

Note that this expression measure is given to you in log base 2 scale. This differs from most of the other expression measure methods.

The tunning factor k will have different meanings if one uses the fast (add-hoc) algorithm or the empirical bayes approach. See Wu et al. (2003)

Value

A matrix of PM intensties.

Author(s)

Rafeal Irizarry & Zhijin Wu

See Also

gcrma.engine2

gcrma.engine2 GCRMA background adjust engine(internal function)

Description

This function adjust for non-specific binding when each array has its own probe affinity information. It takes an AffyBatch object of probe intensities and an AffyBatch of probe affinity, returns one matrix of non-specific binding corrected PM probe intensities.

gcrma.engine2

Arguments

object an AffyBatch. Note: this is an internal function. Optical noise should have

been corrected for.

pmIndex Index of PM probes. This will be computed within the function if left NULL

mmIndex Index of MM probes. This will be computed within the function if left NULL

NCprobe Index of negative control probes. When set as NULL, the MM probes will be

used. These probes are used to estimate parameters of non-specific binding on each array. These will be also used to estimate probe affinity profiles when

affinity.info is not provided.

affinity.info NULL or an AffyBatch containing the affinities in the exprs slot. This object

can be created using the function compute.affinities.

type "fullmodel" for sequence and MM model. "affinities" for sequence information

only. "mm" for using MM without sequence information.

k A tuning factor.

stretch . correction .

GSB. adjust Logical value. If TRUE, probe effects in specific binding will be adjusted.

rho correlation coefficient of log background intensity in a pair of pm/mm probes.

Default=.7

verbose Logical value. If TRUE messages about the progress of the function is printed.

fast Logical value. If TRUE a faster add-hoc algorithm is used.

Details

Note that this expression measure is given to you in log base 2 scale. This differs from most of the other expression measure methods.

The tunning factor k will have different meanings if one uses the fast (add-hoc) algorithm or the empirical bayes approach. See Wu et al. (2003)

Value

A matrix of PM intensties.

Author(s)

Rafeal Irizarry & Zhijin Wu

See Also

gcrma.engine

12 justGCRMA

getCDF	Functions for Automatic Download of Packages

Description

These are internal functions that are called by justGCRMA and GCRMA in order to automatically download and install cdf environments and probe packages.

Usage

```
getCDF(cdfname, lib = .libPaths()[1], verbose = TRUE)
getProbePackage(probepackage, lib = .libPaths()[1], verbose = TRUE)
```

Arguments

cdfname Name of the cdfenv to install.

probepackage Name of the probe package to install.

lib Directory of the R library where packages will be installed.

verbose Output informative comments? Defaults to TRUE

Value

Nothing is returned. These functions are called simply to install environments.

Author(s)

James W. MacDonald, based on getCDFinfo, written by Jeff Gentry.

See Also

getCDFinfo

justGCRMA Compute GCRMA Directly from CEL Files

Description

This function converts CEL files into an ExpressionSet using the robust multi-array average (RMA) expression measure with help of probe sequences.

justGCRMA 13

Usage

```
just.gcrma(..., filenames=character(0),
           phenoData=new("AnnotatedDataFrame"),
           description=NULL.
           notes="", compress=getOption("BioC")$affy$compress.cel,
           normalize=TRUE, bgversion=2, affinity.info=NULL,
           type=c("fullmodel","affinities","mm","constant"),
           k=6*fast+0.5*(1-fast), stretch=1.15*fast+1*(1-fast),
           correction=1, rho=0.7, optical.correct=TRUE,
           verbose=TRUE, fast=TRUE, minimum=1, optimize.by =
           c("speed", "memory"),
           cdfname = NULL, read.verbose = FALSE)
justGCRMA(..., filenames=character(0),
         widget=getOption("BioC")$affy$use.widgets,
         compress=getOption("BioC")$affy$compress.cel,
         celfile.path=getwd(),
         sampleNames=NULL,
         phenoData=NULL,
         description=NULL,
         notes="",
         normalize=TRUE,
         bgversion=2, affinity.info=NULL,
         type=c("fullmodel","affinities","mm","constant"),
         k=6*fast+0.5*(1-fast), stretch=1.15*fast+1*(1-fast),
         correction=1, rho=0.7, optical.correct=TRUE,
         verbose=TRUE, fast=TRUE, minimum=1,
         optimize.by = c("speed", "memory"),
         cdfname = NULL, read.verbose = FALSE)
```

Arguments

... file names separated by comma. filenames file names in a character vector.

widget a logical specifying if widgets should be used.

compress are the CEL files compressed?

phenoData a AnnotatedDataFrame object.

description a MIAME object.

notes notes.

affinity.info NULL or a list of three components: apm,amm and index, for PM probe affinities,

MM probe affinities, the index of probes with known sequence, respectively.

type "fullmodel" for sequence and MM model. "affinities" for sequence information

only. "mm" for using MM without sequence information.

k A tuning factor.

rho correlation coefficient of log background intensity in a pair of pm/mm probes.

Default=.7.

stretch . correction .

14 justGCRMA

normalize Logical value. If TRUE, then normalize data using quantile normalization.

optical.correct

Logical value. If TRUE, then optical background correction is performed.

verbose Logical value. If TRUE, then messages about the progress of the function is

printed.

fast Logical value. If TRUE, then a faster add-hoc algorithm is used.

optimize.by "speed" will use a faster algorithm but more RAM, and "memory" will be slower,

but require less RAM.

bgversion integer value indicating which RMA background to use 1: use background simi-

lar to pure R rma background given in affy version 1.0 - 1.0.2 2: use background

similar to pure R rma background given in affy version 1.1 and above.

minimum .

celfile.path a character denoting the path 'ReadAffy' should look for cel files. sampleNames a character vector of sample names to be used in the 'AffyBatch'.

cdfname Used to specify the name of an alternative cdf package. If set to NULL, the

usual cdf package based on Affymetrix' mappings will be used. Note that the name should not include the 'cdf' on the end, and that the corresponding probe package is also required to be installed. If either package is missing an error will

result.

read.verbose Logical value. If TRUE, then messages will be printed as each celfile is read in.

Details

This method should require much less RAM than the conventional method of first creating an AffyBatch and then running gcrma.

This is a simpler version than gcrma, so some of the arguments available in gcrma are not available here. For example, it is not possible to use the MM probes to estimate background. Instead, the internal NSB estimates are used (which is also the default for gcrma).

Note that this expression measure is given to you in log base 2 scale. This differs from most of the other expression measure methods.

The tuning factor k will have different meanings if one uses the fast (add-hoc) algorithm or the empirical Bayes approach. See Wu et al. (2003)

fast.bkg and mem.bkg are two internal functions.

Value

An ExpressionSet object.

Author(s)

James W. MacDonald

Index

```
* datasets
    affinity.spline.coefs, 2
* internal
    fast.bkg, 6
    getCDF, 12
* manip
    bg.adjust.affinities, 2
    bg.adjust.gcrma, 3
    bg.parameters.ns, 5
    compute.affinities, 5
    gcrma, 8
    gcrma.engine, 9
    gcrma.engine2, 10
    justGCRMA, 12
affinity.spline.coefs, 2, 6
AffyBatch, 4, 8, 11
AnnotatedDataFrame, 13
average.for.PAV(bg.parameters.ns), 5
base.profiles (compute.affinities), 5
bg.adjust.affinities, 2
bg.adjust.constant
        (bg.adjust.affinities), 2
bg.adjust.fullmodel
        (bg.adjust.affinities), 2
bg.adjust.gcrma, 3
bg.adjust.mm(bg.adjust.affinities), 2
bg.adjust.optical
        (bg.adjust.affinities), 2
bg.parameters.ns, 5
check.probes (compute.affinities), 5
compute.affinities, 2, 4, 5, 8, 11
compute.affinities2
        (compute.affinities), 5
compute.affinity.coef
        (compute.affinities), 5
ExpressionSet, 14
fast.bkg, 6
gcrma, 2, 3, 5-7, 8, 14
gcrma.engine, 9
```

```
gcrma.engine2, 10
getCDF, 12
getProbePackage (getCDF), 12
GSB.adj (gcrma), 8
just.gcrma (justGCRMA), 12
justGCRMA, 12
left.sigma (bg.parameters.ns), 5
mem.bkg (fast.bkg), 6
MIAME, 13
PAV (bg.parameters.ns), 5
plotBaseProfiles (compute.affinities), 5
```