

Package ‘PIUMA’

July 2, 2025

Type Package

Title Phenotypes Identification Using Mapper from topological data Analysis

Version 1.5.0

Description The PIUMA package offers a tidy pipeline of Topological Data Analysis frameworks to identify and characterize communities in high and heterogeneous dimensional data.

License GPL-3 + file LICENSE

Encoding UTF-8

LazyData false

biocViews Clustering, GraphAndNetwork, DimensionReduction, Network, Classification

VignetteBuilder knitr

Imports cluster, umap, tsne, kernlab, vegan, dbscan, igraph, scales, Hmisc, patchwork, grDevices, stats, methods, SummarizedExperiment

Suggests BiocStyle, knitr, testthat, rmarkdown

Depends R (>= 4.3), ggplot2

RoxygenNote 7.2.3

URL <https://github.com/BioinfoMonzino/PIUMA>

BugReports <https://github.com/BioinfoMonzino/PIUMA/issues>

git_url <https://git.bioconductor.org/packages/PIUMA>

git_branch devel

git_last_commit a82e3e6

git_last_commit_date 2025-04-15

Repository Bioconductor 3.22

Date/Publication 2025-07-01

Author Mattia Chiesa [aut, cre] (ORCID:

<<https://orcid.org/0000-0001-7427-9954>>),

Arianna Dagliati [aut] (ORCID: <<https://orcid.org/0000-0002-5041-0409>>),

Alessia Gerbasi [aut] (ORCID: <<https://orcid.org/0000-0003-4501-1777>>),

Giuseppe Albi [aut],

Laura Ballarini [aut],

Luca Piacentini [aut] (ORCID: <<https://orcid.org/0000-0003-1022-4481>>)

Maintainer Mattia Chiesa <mattia.chiesa@cardiologicomonzino.it>

Contents

checkNetEntropy	2
checkScaleFreeModel	3
dfToDistance	4
dfToProjection	5
df_test_proj	6
getComp	7
getDfMapper	7
getDistMat	8
getJacc	9
getNodeDataMat	9
getOrigData	10
getOutcome	11
getOutcomeFact	11
getScaledData	12
jaccardMatrix	13
makeTDAobj	14
makeTDAobjFromSE	15
mapperCore	16
PIUMA	17
setComp	17
setDfMapper	18
setDistMat	19
setJacc	19
setNodeDataMat	20
setOrigData	21
setOutcome	21
setOutcomeFact	22
setScaledData	23
tdaDfEnrichment	23
TDAobj-class	24
tda_test_data	25
vascEC_meta	25
vascEC_norm	26

Index	27
--------------	-----------

checkNetEntropy	<i>Compute the Network Entropy</i>
-----------------	------------------------------------

Description

This function computes the average of the entropies for each node of a network.

Usage

```
checkNetEntropy(outcome_vect)
```

Arguments

outcome_vect A vector containing the average outcome values for each node of a network.

Details

The average of the entropies is related to the amount of information stored in the network.

Value

The network entropy using each node of a network.

Author(s)

Mattia Chiesa, Laura Ballarini, Luca Piacentini

See Also

[makeTDAobj](#), [dfToDistance](#), [dfToProjection](#), [mapperCore](#), [jaccardMatrix](#), [tdaDfEnrichment](#)

Examples

```
# use example data:
set.seed(1)
entropy <- checkNetEntropy(round(runif(10),0))
```

checkScaleFreeModel	<i>Assessment of Scale-Free model fitting</i>
---------------------	---

Description

This function assesses the fitting to a scale-free net model.

Usage

```
checkScaleFreeModel(x, showPlot = FALSE)
```

Arguments

x	A TDAobj object, processed by the jaccardMatrix
showPlot	Whether the plot has to be generated. Default: FALSE

Details

The scale-free networks show a high negative correlation between k and $p(k)$.

Value

A list containing:

- the estimated gamma value
- The correlation between the k and the degree distribution $p(k)$.
- The p-value of the correlation between the k and the degree distribution $p(k)$.
- The correlation between the logarithm (base 10) of k and the logarithm (base 10) of the degree distribution $p(k)$.
- The p-value of the correlation between the logarithm (base 10) of k and the logarithm (base 10) of the degree distribution $p(k)$.

Author(s)

Mattia Chiesa, Laura Ballarini, Luca Piacentini

See Also

[makeTDAobj](#), [dfToDistance](#), [dfToProjection](#), [mapperCore](#), [jaccardMatrix](#)

Examples

```
## use example data:
data(tda_test_data)
#netModel <- checkScaleFreeModel(tda_test_data)
```

dfToDistance

Compute the Distance Matrix from TDAobj

Description

This function returns the distance matrix computed by using the Pearson's, Euclidean or Gower distance methods. The distances are computed between the rows of a data.frame in the classical form $n \times m$, where n (rows) are observations and m (columns) are features.

Usage

```
dfToDistance(x, distMethod = c("euclidean", "gower", "pearson"))
```

Arguments

<code>x</code>	A TDAobj object, generated by makeTDAobj Rows (n) and columns (m) should be, respectively, observations and features.
<code>distMethod</code>	The distance method to calculate the distance matrix. "euclidean", "gower" and "pearson" values are allowed. Default: "euclidean".

Value

The starting TDAobj object, in which the computed distance matrix has been added (slot: 'dist_mat')

Author(s)

Mattia Chiesa, Laura Ballarini, Luca Piacentini

See Also

[makeTDAobj](#)

Examples

```
## use example data:
data(tda_test_data)
dfDist <- dfToDistance(tda_test_data, "euclidean")
```

dfToProjection

*Data projection using a Dimensionality Reduction Method***Description**

This function performs the transformation of data from a high dimensional space into a low dimensional space, wrapping 6 well-known reduction methods; i.e., PCA, KPCA, t-SNE, UMAP, MDS, and Isomap. In the topological data analysis, the identified components are commonly used as lenses.

Usage

```
dfToProjection(
  x,
  method = c("PCA", "UMAP", "TSNE", "MDS", "KPCA", "ISOMAP"),
  nComp = 2,
  centerPCA = FALSE,
  scalePCA = FALSE,
  umapNNeigh = 15,
  umapMinDist = 0.1,
  tsnePerpl = 30,
  tsneMaxIter = 300,
  kpcaKernel = c("rbfdot", "laplacedot", "polydot", "tanhdot", "besseldot", "anovadot",
    "vanilladot", "splinedot"),
  kpcaSigma = 0.1,
  kpcaDegree = 1,
  isomNNeigh = 5,
  showPlot = FALSE,
  vectColor = NULL
)
```

Arguments

x	A TDAobj object, generated by makeTDAobj
method	Name of the dimensionality reduction method to use. "PCA", "UMAP", "TSNE", "MDS", "KPCA" and "isomap" values are allowed. Default is: "PCA".
nComp	The number of components to be computed. Default: 2
centerPCA	Whether the data should be centered before PCA. Default: TRUE
scalePCA	Whether the data should be scaled before PCA. Default: TRUE
umapNNeigh	The number of neighbors for UMAP. Default: 15
umapMinDist	The minimum distance between points for UMAP. Default: 0.1
tsnePerpl	Perplexity argument of t-SNE. Default: 30
tsneMaxIter	The maximum number of iterations for t-SNE. Default: 300
kpcaKernel	The type of kernel for kPCA. "rbfdot", "laplacedot", "polydot", "tanhdot", "besseldot", "anovadot", "vanilladot" and "splinedot" are allowed. Default: "polydot".
kpcaSigma	The 'sigma' argument for kPCA. Default: 0.1.
kpcaDegree	The 'degree' argument for kPCA. Default: 1.

isomNNeigh	The number of neighbors for Isomap. Default: 5.
showPlot	Whether the scatter plot of the first two principal components should be shown. Default: TRUE.
vectColor	Vector containing the variable to color the scatter plot Default: NULL.

Value

The starting TDAobj object, in which the principal components of projected data have been added (slot: 'comp')

Author(s)

Mattia Chiesa, Laura Ballarini, Luca Piacentini

See Also

[makeTDAobj](#), [dfToDistance](#)

Examples

```
# use example data:
data(tda_test_data)
set.seed(1)
cmp <- dfToProjection(tda_test_data, "PCA", nComp=2)
```

df_test_proj	<i>A dataset to test the dfToProjection and dfToDistance functions of PIUMA package.</i>
--------------	--

Description

A dataset to test the [dfToProjection](#) and [dfToDistance](#) functions of PIUMA package.

Usage

```
df_test_proj
```

Format

A data.frame containing 15 rows (cells) and 15 columns (genes)

Value

An example dataset for PIUMA package

`getComp`*Getter method for the 'comp' slot of a TDAobj object.*

Description

The method to get data from the comp slot

Usage

```
getComp(x)
```

```
## S4 method for signature 'TDAobj'  
getComp(x)
```

Arguments

x a TDAobj object

Value

a data.frame with the comp data

Author(s)

Mattia Chiesa

Examples

```
data(tda_test_data)
```

`getDfMapper`*Getter method for the 'dfMapper' slot of a TDAobj object.*

Description

The method to get data from the dfMapper slot

Usage

```
getDfMapper(x)
```

```
## S4 method for signature 'TDAobj'  
getDfMapper(x)
```

Arguments

x a TDAobj object

Value

a data.frame with the dfMapper data

Author(s)

Mattia Chiesa

Examples

```
data(tda_test_data)
ex_out <- getDfMapper(tda_test_data)
```

getDistMat

Getter method for the 'dist_mat' slot of a TDAobj object.

Description

The method to get data from the dist_mat slot

Usage

```
getDistMat(x)

## S4 method for signature 'TDAobj'
getDistMat(x)
```

Arguments

x a TDAobj object

Value

a data.frame with the dist_mat data

Author(s)

Mattia Chiesa

Examples

```
data(tda_test_data)
ex_out <- getDistMat(tda_test_data)
```

`getJacc`*Getter method for the 'jacc' slot of a TDAobj object.*

Description

The method to get data from the jacc slot

Usage

```
getJacc(x)
```

```
## S4 method for signature 'TDAobj'  
getJacc(x)
```

Arguments

x a TDAobj object

Value

a matrix with the jacc data

Author(s)

Mattia Chiesa

Examples

```
data(tda_test_data)  
ex_out <- getJacc(tda_test_data)
```

`getNodeDataMat`*Getter method for the 'node_data_mat' slot of a TDAobj object.*

Description

The method to get data from the node_data_mat slot

Usage

```
getNodeDataMat(x)
```

```
## S4 method for signature 'TDAobj'  
getNodeDataMat(x)
```

Arguments

x a TDAobj object

Value

a data.frame with the node_data_mat data

Author(s)

Mattia Chiesa

Examples

```
data(tda_test_data)
ex_out <- getNodeDataMat(tda_test_data)
```

getOrigData

Getter method for the 'orig_data' slot of a TDAobj object.

Description

The method to get data from the orig_data slot

Usage

```
getOrigData(x)

## S4 method for signature 'TDAobj'
getOrigData(x)
```

Arguments

x a TDAobj object

Value

a data.frame with the original data

Author(s)

Mattia Chiesa

Examples

```
data(tda_test_data)
ex_out <- getOrigData(tda_test_data)
```

getOutcome	Getter method for the 'outcome' slot of a TDAobj object.
------------	--

Description

The method to get data from the outcome slot

Usage

```
getOutcome(x)

## S4 method for signature 'TDAobj'
getOutcome(x)
```

Arguments

x a TDAobj object

Value

a data.frame with the outcome data

Author(s)

Mattia Chiesa

Examples

```
data(tda_test_data)
ex_out <- getOutcome(tda_test_data)
```

getOutcomeFact	Getter method for the 'outcomeFact' slot of a TDAobj object.
----------------	--

Description

The method to get data from the outcomeFact slot

Usage

```
getOutcomeFact(x)

## S4 method for signature 'TDAobj'
getOutcomeFact(x)
```

Arguments

x a TDAobj object

Value

a data.frame with the outcomeFact data

Author(s)

Mattia Chiesa

Examples

```
data(tda_test_data)
ex_out <- getOutcomeFact(tda_test_data)
```

getScaledData

Getter method for the 'scaled_data' slot of a TDAobj object.

Description

The method to get data from the scaled_data slot

Usage

```
getScaledData(x)

## S4 method for signature 'TDAobj'
getScaledData(x)
```

Arguments

x a TDAobj object

Value

a data.frame with the scaled data

Author(s)

Mattia Chiesa

Examples

```
data(tda_test_data)
ex_out <- getScaledData(tda_test_data)
```

jaccardMatrix*Compute the Matrix of Jaccard Indexes*

Description

This function computes the Jaccard index for each pair of nodes contained in TDAobj, generated by the [mapperCore](#) function. The resulting data.frame can be used to represent data as a network, for instance, in Cytoscape

Usage

```
jaccardMatrix(x)
```

Arguments

x A TDAobj object, processed by the [mapperCore](#) function.

Details

The Jaccard index measures the similarity of two nodes A and B. It ranges from 0 to 1. If A and B share no members, their Jaccard index would be 0 (= NA). If A and B share all members, their Jaccard index would be 1. Hence, the higher the index, the more similar the two nodes. If the Jaccard index between A and B is different from NA, it means that an edge exists between A and B. The output matrix of Jaccard indexes can be used as an adjacency matrix. The resulting data.frame can be used to represent data as a network, for instance, in Cytoscape.

Value

The starting TDAobj object, in which the matrix of Jaccard indexes, calculated comparing each node of the 'dfMapper' slot, has been added (slot: 'jacc')

Author(s)

Mattia Chiesa, Laura Ballarini, Luca Piacentini

See Also

[makeTDAobj](#), [dfToDistance](#), [dfToProjection](#), [mapperCore](#)

Examples

```
## use example data:
data(tda_test_data)
jacc_mat <- jaccardMatrix(tda_test_data)
```

makeTDAobj

*Import data and generate the TDAobj object***Description**

This function import a data.frame and create the object to store all data needed for TDA analysis. In addition, some preliminary preprocess steps are performed; specifically, outcomes variables data will be separated the rest of dataset. The remaining dataset will be also re-scaled (0-1)

Usage

```
makeTDAobj(df, outcomes)
```

Arguments

df	A data.frame representing a dataset in the classical $n \times m$ form. Rows (n) and columns (m) should be, respectively, observations and features.
outcomes	A string or vector of string containing the name of variables that have to be considered 'outcomes'

Value

A TDA object containing:

- orig_data A data.frame of original data (without outcomes)
- scaled_data A data.frame of re-scaled data (without outcomes)
- outcomeFact A data.frame of original outcomes
- outcome A data.frame of original outcomes converted as numeric
- comp A data.frame containing the components of projected data
- dist_mat A data.frame containing the computed distance matrix
- dfMapper A data.frame containing the nodes, with their elements, identified by TDA
- jacc A matrix of Jaccard indexes between each pair of dfMapper nodes
- node_data_mat A data.frame with the node size and the average value

Author(s)

Mattia Chiesa, Laura Ballarini, Luca Piacentini

Examples

```
## use example data:
data("vascEC_meta")
data("vascEC_norm")
df <- cbind(vascEC_meta, vascEC_norm)
res <- makeTDAobj(df, "zone")
```

makeTDAobjFromSE	<i>Import SummarizedExperiment data and generate the TDAobj object</i>
------------------	--

Description

This function import a SummarizedExperiment object and create the object to store all data needed for TDA analysis. In addition, some preliminary preprocess steps are performed; specifically, outcomes variables data will be separated the rest of dataset. The remaining dataset will be also re-scaled (0-1)

Usage

```
makeTDAobjFromSE(SE, outcomes)
```

Arguments

SE	A SummarizedExperiment object
outcomes	A string or vector of string containing the name of variables that have to be considered 'outcomes'

Value

A TDA object containing:

- orig_data A data.frame of original data (without outcomes)
- scaled_data A data.frame of re-scaled data (without outcomes)
- outcomeFact A data.frame of original outcomes
- outcome A data.frame of original outcomes converted as numeric
- comp A data.frame containing the components of projected data
- dist_mat A data.frame containing the computed distance matrix
- dfMapper A data.frame containing the nodes, with their elements, identified by TDA
- jacc A matrix of Jaccard indexes between each pair of dfMapper nodes
- node_data_mat A data.frame with the node size and the average value

Author(s)

Mattia Chiesa, Laura Ballarini, Luca Piacentini

Examples

```
## use example data:
data("vascEC_meta")
data("vascEC_norm")
suppressMessages(library(SummarizedExperiment))
dataSE <- SummarizedExperiment(assays=as.matrix(t(vascEC_norm)),
                              colData=as.data.frame(vascEC_meta))
res <- makeTDAobjFromSE(dataSE, "zone")
```

mapperCore

Implement the TDA Mapper algorithm on TDAobj

Description

This is a comprehensive function permitting to perform the core TDA Mapper algorithm with 2D lenses. It allow setting several types of clustering methods.

Usage

```
mapperCore(
  x,
  nBins = 15,
  overlap = 0.4,
  mClustNode = 2,
  remEmptyNode = TRUE,
  clustMeth = c("kmeans", "HR", "DBSCAN", "OPTICS"),
  HRMethod = c("average", "complete")
)
```

Arguments

x	A TDAobj object, processed by the dfToDistance and dfToProjection functions.
nBins	The number of bins (i.e. the resolution of the cover). Default: 15.
overlap	The overlap between bins (i.e.the gain of the cover). Default: 0.4.
mClustNode	The number of clusters in each overlapping bin. Default: 2
remEmptyNode	A logical value to remove or not the empty nodes from the resulting data.frame. Default: TRUE.
clustMeth	The clustering algorithm."HR", "kmeans", "DBSCAN", and "OPTICS" are allowed. Default: "kmeans".
HRMethod	The name of the linkage criterion (when clustMeth="HR"). "average" and "complete" values are allowed. Default: "average".

Value

The starting TDAobj object, in which the result of mapper algorithm (inferred nodes with their elements) has been added (slot: 'dfMapper')

A data.frame containing the clusters, with their elements, identified by TDA .

Author(s)

Mattia Chiesa, Laura Ballarini, Luca Piacentini

See Also

[makeTDAobj](#), [dfToDistance](#), [dfToProjection](#)

Examples

```
# use example data:
data(tda_test_data)
set.seed(1)
dfMapper <- mapperCore(tda_test_data, nBins=5, overlap=0.5,
mClustNode=2, clustMeth="kmeans")
```

PIUMA

PIUMA: Phenotypes Identification Using Mapper from topological data Analysis

Description

The application of unsupervised learning methodologies could help the identification of specific phenotypes in huge heterogeneous cohorts, such as clinical or -omics data. Among them, the Topological Data Analysis (TDA) is a rapidly growing field that combines concepts from algebraic topology and computational geometry to analyze and extract meaningful information from complex and high-dimensional data sets. Moreover, TDA is a robust and effective methodology, able to preserve the intrinsic characteristics of data and the mutual relations among observations, depicting complex data in a graph-based representation. Indeed, building topological models as networks, TDA allows complex diseases to be inspected in a continuous space, where subjects can fluctuate over the graph, sharing, at the same time, more than one adjacent node of the network. Overall, TDA offers a powerful set of tools to capture the underlying topological features of data, revealing essential patterns and relationships that might be hidden from traditional statistical techniques. The PIUMA package (Phenotypes Identification Using Mapper from topological data Analysis) allows implementing all the main steps of a Topological Data Analysis. PIUMA is the italian word meaning 'feather'.

Details

See the package vignette, by typing `vignette("PIUMA")` to discover all the functions.

Author(s)

Mattia Chiesa, Laura Ballarini, Luca Piacentini

setComp

Setter method for the 'comp' slot of a TDAobj object.

Description

The method to set the comp slot

Usage

```
setComp(x, y)

## S4 method for signature 'TDAobj'
setComp(x, y)
```

Arguments

x a TDAobj object
y a data.frame with the comp data

Value

a TDAobj object

Author(s)

Mattia Chiesa

Examples

```
data(tda_test_data)
```

setDfMapper	<i>Setter method for the 'dfMapper' slot of a TDAobj object.</i>
-------------	--

Description

The method to set the dfMapper slot

Usage

```
setDfMapper(x, y)  
  
## S4 method for signature 'TDAobj'  
setDfMapper(x, y)
```

Arguments

x a TDAobj object
y a data.frame with the dfMapper data

Value

a TDAobj object

Author(s)

Mattia Chiesa

Examples

```
data(tda_test_data)
```

setDistMat	<i>Setter method for the 'dist_mat' slot of a TDAobj object.</i>
------------	--

Description

The method to set the dist_mat slot

Usage

```
setDistMat(x, y)
```

```
## S4 method for signature 'TDAobj'
setDistMat(x, y)
```

Arguments

x	a TDAobj object
y	a data.frame with the dist_mat data

Value

a TDAobj object

Author(s)

Mattia Chiesa

Examples

```
data(tda_test_data)
```

setJacc	<i>Setter method for the 'jacc' slot of a TDAobj object.</i>
---------	--

Description

The method to set the jacc slot

Usage

```
setJacc(x, y)
```

```
## S4 method for signature 'TDAobj'
setJacc(x, y)
```

Arguments

x	a TDAobj object
y	a matrix with the jacc data

Value

a TDAobj object

Author(s)

Mattia Chiesa

Examples

```
data(tda_test_data)
```

setNodeDataMat

Setter method for the 'node_data_mat' slot of a TDAobj object.

Description

The method to set the node_data_mat slot

Usage

```
setNodeDataMat(x, y)
```

```
## S4 method for signature 'TDAobj'  
setNodeDataMat(x, y)
```

Arguments

x	a TDAobj object
y	a data.frame with the node_data_mat data

Value

a TDAobj object

Author(s)

Mattia Chiesa

Examples

```
data(tda_test_data)
```

setOrigData	<i>Setter method for the 'orig_data' slot of a TDAobj object.</i>
-------------	---

Description

The method to set the orig_data slot

Usage

```
setOrigData(x, y)
```

```
## S4 method for signature 'TDAobj'  
setOrigData(x, y)
```

Arguments

x	a TDAobj object
y	a data.frame with the original data

Value

a TDAobj object

Author(s)

Mattia Chiesa

Examples

```
data(tda_test_data)
```

setOutcome	<i>Setter method for the 'outcome' slot of a TDAobj object.</i>
------------	---

Description

The method to set the outcome slot

Usage

```
setOutcome(x, y)
```

```
## S4 method for signature 'TDAobj'  
setOutcome(x, y)
```

Arguments

x	a TDAobj object
y	a data.frame with the outcome data

Value

a TDAobj object

Author(s)

Mattia Chiesa

Examples

```
data(tda_test_data)
```

setOutcomeFact

Setter method for the 'outcomeFact' slot of a TDAobj object.

Description

The method to set the outcomeFact slot

Usage

```
setOutcomeFact(x, y)

## S4 method for signature 'TDAobj'
setOutcomeFact(x, y)
```

Arguments

x	a TDAobj object
y	a data.frame with the outcomeFact data

Value

a TDAobj object

Author(s)

Mattia Chiesa

Examples

```
data(tda_test_data)
```

setScaledData	<i>Setter method for the 'scaled_data' slot of a TDAobj object.</i>
---------------	---

Description

The method to set the scaled_data slot

Usage

```
setScaledData(x, y)

## S4 method for signature 'TDAobj'
setScaledData(x, y)
```

Arguments

x	a TDAobj object
y	a data.frame with the scaled data

Value

a TDAobj object

Author(s)

Mattia Chiesa

Examples

```
data(tda_test_data)
```

tdaDfEnrichment	<i>Add information to TDAobj</i>
-----------------	----------------------------------

Description

This function computes the average value of additional features provided by the user and calculate the size for each node of 'dfMapper' slot

Usage

```
tdaDfEnrichment(x, df)
```

Arguments

x	A TDAobj object, processed by the mapperCore function.
df	A data.frame with scaled values in the classical n x m form: rows (n) and columns (m) must be observations and features, respectively.

Value

The starting TDAobj object, in which the a data.frame with additional information for each node has been added (slot: 'node_data_mat')

Author(s)

Mattia Chiesa, Laura Ballarini, Luca Piacentini

See Also

[makeTDAobj](#), [dfToDistance](#), [dfToProjection](#), [mapperCore](#), [jaccardMatrix](#)

Examples

```
## use example data:
data(tda_test_data)
data(df_test_proj)
enrich_mat_tda <- tdaDfEnrichment(tda_test_data, df_test_proj)
```

TDAobj-class

The object 'TDAobj'

Description

The TDA object for storing TDA data

Value

TDAobj class `showClass("TDAobj")`

Slots

`orig_data` A data.frame of original data (without outcomes)
`scaled_data` A data.frame of re-scaled data (without outcomes)
`outcomeFact` A data.frame of original outcomes
`outcome` A data.frame of original outcomes converted as numeric
`comp` A data.frame containing the components of projected data
`dist_mat` A data.frame containing the computed distance matrix
`dfMapper` A data.frame containing the nodes, with their elements, identified by TDA
`jacc` A matrix of Jaccard indexes between each pair of dfMapper nodes
`node_data_mat` A data.frame with the node size and the average value of each feature

tda_test_data	<i>A TDAobj to test the PIUMA package.</i>
---------------	--

Description

A TDAobj to test the PIUMA package.

Usage

```
tda_test_data
```

Format

A TDAobj with data in all slots

Value

An example dataset for PIUMA package

vascEC_meta	<i>Example datasets for PIUMA package</i>
-------------	---

Description

We tested PIUMA on a subset of the single-cell RNA Sequencing dataset (GSE:GSE193346 generated and published by Feng et al. (2022) on Nature Communication to demonstrate that distinct transcriptional profiles are present in specific cell types of each heart chambers, which were attributed to have roles in cardiac development. In this tutorial, our aim will be to exploit PIUMA for identifying sub-population of vascular endothelial cells, which can be associated with specific heart developmental stages. The original dataset consisted of three layers of heterogeneity: cell type, stage and zone (i.e., heart chamber). Our testing dataset was obtained by subsetting vascular endothelial cells (cell type) by Seurat object, extracting raw counts and metadata. Thus, we filtered low expressed genes and normalized data by DaMiRseq

Usage

```
vascEC_meta
```

Format

A dataframe containing 1180 rows (cells) and 2 columns (outcomes)

Value

An example dataset for PIUMA package

vascEC_norm

We tested PIUMA on a subset of the single-cell RNA Sequencing dataset (GSE:GSE193346 generated and published by Feng et al. (2022) on Nature Communication to demonstrate that distinct transcriptional profiles are present in specific cell types of each heart chambers, which were attributed to have roles in cardiac development. In this tutorial, our aim will be to exploit PIUMA for identifying sub-population of vascular endothelial cells, which can be associated with specific heart developmental stages. The original dataset consisted of three layers of heterogeneity: cell type, stage and zone (i.e., heart chamber). Our testing dataset was obtained by subsetting vascular endothelial cells (cell type) by Seurat object, extracting raw counts and metadata. Thus, we filtered low expressed genes and normalized data by DaMiRseq

Description

We tested PIUMA on a subset of the single-cell RNA Sequencing dataset (GSE:GSE193346 generated and published by Feng et al. (2022) on Nature Communication to demonstrate that distinct transcriptional profiles are present in specific cell types of each heart chambers, which were attributed to have roles in cardiac development. In this tutorial, our aim will be to exploit PIUMA for identifying sub-population of vascular endothelial cells, which can be associated with specific heart developmental stages. The original dataset consisted of three layers of heterogeneity: cell type, stage and zone (i.e., heart chamber). Our testing dataset was obtained by subsetting vascular endothelial cells (cell type) by Seurat object, extracting raw counts and metadata. Thus, we filtered low expressed genes and normalized data by DaMiRseq

Usage

vascEC_norm

Format

A matrix containing 1180 rows (cells) and 838 columns (genes)

Value

An example dataset for PIUMA package

Index

* datasets

df_test_proj, [6](#)
tda_test_data, [25](#)
vascEC_meta, [25](#)
vascEC_norm, [26](#)

* package

PIUMA, [17](#)

checkNetEntropy, [2](#)
checkScaleFreeModel, [3](#)

df_test_proj, [6](#)
dfToDistance, [3](#), [4](#), [4](#), [6](#), [13](#), [16](#), [24](#)
dfToProjection, [3](#), [4](#), [5](#), [6](#), [13](#), [16](#), [24](#)

getComp, [7](#)
getComp, PIUMA-getComp (getComp), [7](#)
getComp, TDAobj-method (getComp), [7](#)
getDfMapper, [7](#)
getDfMapper, PIUMA-getDfMapper
(getDfMapper), [7](#)
getDfMapper, TDAobj-method
(getDfMapper), [7](#)
getDistMat, [8](#)
getDistMat, PIUMA-getDistMat
(getDistMat), [8](#)
getDistMat, TDAobj-method (getDistMat), [8](#)
getJacc, [9](#)
getJacc, PIUMA-getJacc (getJacc), [9](#)
getJacc, TDAobj-method (getJacc), [9](#)
getNodeDataMat, [9](#)
getNodeDataMat, PIUMA-getNodeDataMat
(getNodeDataMat), [9](#)
getNodeDataMat, TDAobj-method
(getNodeDataMat), [9](#)
getOrigData, [10](#)
getOrigData, PIUMA-getOrigData
(getOrigData), [10](#)
getOrigData, TDAobj-method
(getOrigData), [10](#)
getOutcome, [11](#)
getOutcome, PIUMA-getOutcome
(getOutcome), [11](#)

getOutcome, TDAobj-method (getOutcome),
[11](#)
getOutcomeFact, [11](#)
getOutcomeFact, PIUMA-getOutcomeFact
(getOutcomeFact), [11](#)
getOutcomeFact, TDAobj-method
(getOutcomeFact), [11](#)
getScaledData, [12](#)
getScaledData, PIUMA-getScaledData
(getScaledData), [12](#)
getScaledData, TDAobj-method
(getScaledData), [12](#)

jaccardMatrix, [3](#), [4](#), [13](#), [24](#)

makeTDAobj, [3–6](#), [13](#), [14](#), [16](#), [24](#)
makeTDAobjFromSE, [15](#)
mapperCore, [3](#), [4](#), [13](#), [16](#), [23](#), [24](#)

PIUMA, [17](#)

setComp, [17](#)
setComp, PIUMA-setComp (setComp), [17](#)
setComp, TDAobj-method (setComp), [17](#)
setDfMapper, [18](#)
setDfMapper, PIUMA-setDfMapper
(setDfMapper), [18](#)
setDfMapper, TDAobj-method
(setDfMapper), [18](#)
setDistMat, [19](#)
setDistMat, PIUMA-setDistMat
(setDistMat), [19](#)
setDistMat, TDAobj-method (setDistMat),
[19](#)
setJacc, [19](#)
setJacc, PIUMA-setJacc (setJacc), [19](#)
setJacc, TDAobj-method (setJacc), [19](#)
setNodeDataMat, [20](#)
setNodeDataMat, PIUMA-setNodeDataMat
(setNodeDataMat), [20](#)
setNodeDataMat, TDAobj-method
(setNodeDataMat), [20](#)
setOrigData, [21](#)
setOrigData, PIUMA-setOrigData
(setOrigData), [21](#)

setOrigData, TDAobj-method
 (setOrigData), [21](#)
setOutcome, [21](#)
setOutcome, PIUMA-setOutcome
 (setOutcome), [21](#)
setOutcome, TDAobj-method (setOutcome),
 [21](#)
setOutcomeFact, [22](#)
setOutcomeFact, PIUMA-setOutcomeFact
 (setOutcomeFact), [22](#)
setOutcomeFact, TDAobj-method
 (setOutcomeFact), [22](#)
setScaledData, [23](#)
setScaledData, PIUMA-setScaledData
 (setScaledData), [23](#)
setScaledData, TDAobj-method
 (setScaledData), [23](#)

tda_test_data, [25](#)
tdaDfEnrichment, [3](#), [23](#)
TDAobj-class, [24](#)

vascEC_meta, [25](#)
vascEC_norm, [26](#)