

Package ‘RPA’

July 5, 2025

Type Package

Title RPA: Robust Probabilistic Averaging for probe-level analysis

Encoding UTF-8

Version 1.65.0

Date 2023-04-07

Depends R (>= 3.1.1), affy, BiocGenerics, BiocStyle, methods,
rmarkdown

Imports phyloseq

Suggests knitr, parallel

biocViews GeneExpression, Microarray, Preprocessing, QualityControl

Description Probabilistic analysis of probe reliability and
differential gene expression on short oligonucleotide
arrays.

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URL <https://github.com/antagomir/RPA>

BugReports <https://github.com/antagomir/RPA>

VignetteBuilder knitr

RoxygenNote 7.2.3

git_url <https://git.bioconductor.org/packages/RPA>

git_branch devel

git_last_commit 389f93f

git_last_commit_date 2025-04-15

Repository Bioconductor 3.22

Date/Publication 2025-07-04

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RPA-package

RPA: probabilistic analysis of probe reliability and gene expression

Description

Brief summary of the RPA package

Details

Package: RPA
Type: Package
Version: See sessionInfo() or DESCRIPTION file
Date: 2008-2016
License: FreeBSD
LazyLoad: yes

Author(s)

Leo Lahti <leo.lahti@iki.fi>

References

See citation("RPA")

Examples

#

calculate.rpa	<i>RPA with HITChip</i>
---------------	-------------------------

Description

Fit RPA for HITChip.

Usage

```
calculate.rpa(level, phylo, oligo.data)
```

Arguments

level	level
phylo	phylo
oligo.data	oligo.data

Value

RPA preprocessed data

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

See citation("microbiome")

```
collect.hyperparameters
```

collect.hyperparameters

Description

Collect probe-level parameters during online-learning from the batch files.

Usage

```
collect.hyperparameters(  
  batches,  
  unique.run.identifier,  
  save.batches.dir,  
  save.batches,  
  verbose = TRUE  
)
```

Arguments

batches	batch list
unique.run.identifier	Batch file identifier string
save.batches.dir	Batch file directory
save.batches	Logical. Determines whether batches are available.
verbose	verbose

Author(s)

Leo Lahti <leo.lahti@iki.fi>

References

See citation("RPA")

Examples

```
# hpe <- collect.hyperparameters(batches, unique.run.identifier, save.batches.dir, save.batches)
```

d.update.fast	<i>Fast d update</i>
---------------	----------------------

Description

Computes weighted average over the probes, weighted by their inverse probe-specific variances.

Usage

```
d.update.fast(St, s2)
```

Arguments

St	probes x samples data matrix
s2	variances for the probes

Details

Returns summarized probeset-level weighted average

Author(s)

Leo Lahti <leo.lahti@iki.fi>

References

See citation("RPA")

Examples

```
#
```

estimate.affinities	<i>estimate.affinities</i>
---------------------	----------------------------

Description

Probe affinity estimation. Estimates probe-specific affinity parameters.

Usage

```
estimate.affinities(dat, a)
```

Arguments

dat	Input data set: probes x samples.
a	Estimated expression signal from RPA model.

Details

To estimate means in the original data domain let us assume that each probe-level observation x is of the following form: $x = d + v + \text{noise}$, where x and d are vectors over samples, v is a scalar (vector with identical elements) noise is Gaussian with zero mean and probe-specific variance parameters τ^2 . Then the parameter μ will indicate how much probe-level observation deviates from the estimated signal shape d . This deviation is further decomposed as $\mu = \mu.\text{real} + \mu.\text{probe}$, where $\mu.\text{real}$ describes the 'real' signal level, common for all probes $\mu.\text{probe}$ describes probe affinity effect. Let us now assume that $\mu.\text{probe} \sim N(0, \sigma.\text{probe})$. This encodes the assumption that in general the affinity effect of each probe tends to be close to zero. Then we just calculate ML estimates of $\mu.\text{real}$ and $\mu.\text{probe}$ based on particular assumptions. Note that this part of the algorithm has not been defined in full probabilistic terms yet, just calculating the point estimates. Note that while τ^2 in RPA measures stochastic noise, and NOT the affinity effect, we use it here as a heuristic solution to weigh the probes according to how much they contribute to the overall signal shape. Intuitively, probes that have little effect on the signal shape (i.e. are very noisy and likely to be contaminated by many unrelated signals) should also contribute less to the absolute signal estimate. If no other prior information is available, using stochastic parameters τ^2 to determine probe weights is likely to work better than simple averaging of the probes without weights. Also in this case the probe affinities sum close to zero but there is some flexibility, and more noisy probes can be downweighted.

Value

A vector with probe-specific affinities.

Author(s)

Leo Lahti <leo.lahti@iki.fi>

References

See citation("RPA")

See Also

rpa.fit

Examples

```
# mu <- estimate.affinities(dat, a)
```

```
estimate.hyperparameters
      estimate.hyperparameters
```

Description

Hyperparameter estimation.

Usage

```
estimate.hyperparameters(
  sets = NULL,
  probe.parameters = list(alpha = 2, beta = 1),
  batches,
  cdf = NULL,
  bg.method = "rma",
  epsilon = 0.01,
  load.batches = FALSE,
  save.hyperparameter.batches = FALSE,
  mc.cores = 1,
  verbose = TRUE,
  normalization.method = "quantiles",
  save.batches.dir = ".",
  unique.run.identifier = NULL,
  set.inds = set.inds
)
```

Arguments

<code>sets</code>	Probesets to handle. All probesets by default.
<code>probe.parameters</code>	User-defined priors. May also include <code>quantile.basis</code>
<code>batches</code>	Data batches for online learning
<code>cdf</code>	CDF probeset definition file
<code>bg.method</code>	Background correction method
<code>epsilon</code>	Convergence parameter
<code>load.batches</code>	Logical. Load preprocessed data whose identifiers are picked from <code>names(batches)</code> . Assuming that the same batch list (<code>batches</code>) was used to create the files in <code>on-line.quantiles</code> function.
<code>save.hyperparameter.batches</code>	Save hyperparameters for each batch into files using the identifiers with batch name with <code>-hyper.RData</code> suffix.
<code>mc.cores</code>	Number of cores for parallel computation
<code>verbose</code>	Print progress information
<code>normalization.method</code>	Normalization method
<code>save.batches.dir</code>	Specify the output directory for temporary batch saves.
<code>unique.run.identifier</code>	Define identifier for this run for naming the temporary batch files. By default, a random id is generated.
<code>set.inds</code>	Probeset indices

Value

`alpha`: Hyperparameter alpha (same for all probesets); `betas`: Hyperparameter beta (probe-specific); `variances`: Probe-specific variances (`beta/alpha`)

Author(s)

Leo Lahti <leo.lahti@iki.fi>

References

See citation("RPA")

Examples

```
#
```

frpa

frpa

Description

Frozen-RPA preprocessing using precalculated probe parameters.

Usage

```
frpa(
  abatch = NULL,
  probe.parameters = NULL,
  verbose = FALSE,
  cdf = NULL,
  cel.files = NULL,
  cel.path = NULL,
  mc.cores = 1,
  summarize.with.affinities = FALSE
)
```

Arguments

abatch An AffyBatch object.

probe.parameters

A list with tau2 (probe variance), quantile.basis (basis for quantile normalization in log2 domain), and optionally affinity (probe affinities). The probe.parameters\$tau2 and probe.parameters\$affinity are lists, each element corresponding to a probe-set and containing a parameter vector over the probes. The quantile.basis is a vector over the probes, the probes need to be listed in the same order as in tau2 and affinity. probe.parameters can be optionally provided as a data frame.

verbose Print progress information during computation.

cdf Specify an alternative CDF environment. Default: none.

cel.files List of CEL files to preprocess.

cel.path Path to CEL file directory.

mc.cores Number of cores for parallelized processing.

summarize.with.affinities

Use affinity estimates in probe summarization step. Default: FALSE.

Details

fRPA function to preprocess Affymetrix CEL files with RPA using precalculated (frozen) probe parameters.

Value

Preprocessed expression matrix in expressionSet format

Author(s)

Leo Lahti <leo.lahti@iki.fi>

References

See citation("RPA")

See Also

rpa, AffyBatch, ExpressionSet

Examples

```
# eset <- frpa(abatch, probe.parameters)
```

get.batches

get.batches Split data into batches

Description

get.batches Split data into batches

Usage

```
get.batches(items, batch.size = NULL, shuffle = FALSE)
```

Arguments

items	A vector of items to be splitted into batches.
batch.size	Batch size. The last batch may contain less elements than the other batches which have batch.size elements each.
shuffle	Split the elements randomly in the batches.

Value

A list. Each element corresponds to one batch and contains a vector listing the elements in that batch.

Author(s)

Leo Lahti <leo.lahti@iki.fi>

References

See citation("RPA")

Examples

```
#
```

get.probe.matrix	<i>get.probe.matrix</i>
------------------	-------------------------

Description

Get probe matrix.

Usage

```
get.probe.matrix(
  cels,
  cdf = NULL,
  quantile.basis,
  bg.method = "rma",
  normalization.method = "quantiles",
  batch = NULL,
  verbose = TRUE
)
```

Arguments

cels	List of CEL files to preprocess
cdf	Specify an alternative CDF environment
quantile.basis	Pre-calculated basis for quantile normalization in log2 domain
bg.method	Specify background correction method. See bgcorrect.methods() for options.
normalization.method	normalization method
batch	batch
verbose	Print progress information during computation

Details

Returns background-corrected, quantile normalized log2 probes x samples matrix

Author(s)

Leo Lahti <leo.lahti@iki.fi>

References

See citation("RPA")

Examples

```
#
```

get.probe.parameters	<i>get.probe.parameters</i>
----------------------	-----------------------------

Description

Get probe-level hyperparameter from batch files

Usage

```
get.probe.parameters(
  affinities,
  unique.run.identifier,
  save.batches.dir = ".",
  mode = "list"
)
```

Arguments

affinities	probe affinities
unique.run.identifier	Batch file identifier string
save.batches.dir	Batch file directory
mode	"list" or "table"

Author(s)

Leo Lahti <leo.lahti@iki.fi>

References

See citation("RPA")

Examples

```
# df <- get.probe.parameters(unique.run.identifier, save.batches.dir = ".", mode = "list")
```

get.probeset	<i>Get probeset</i>
--------------	---------------------

Description

Get probeset matrix.

Usage

```
get.probeset(name, level, taxonomy, probedata, log10 = TRUE)
```

Arguments

name	name
level	taxonomic level
taxonomy	taxonomy
probedata	oligos vs. samples preprocessed data matrix; absolute scale
log10	Logical. Logarithmize the data TRUE/FALSE

Value

probeset data matrix

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

See citation('microbiome')

Examples

```
#taxonomy <- GetPhylogeny('HITChip', 'filtered')
#data.dir <- system.file("extdata", package = "microbiome")
#probedata <- read_hitchip(data.dir, "rpa")$probedata
#ps <- get.probeset('Akkermansia', 'L2', taxonomy, probedata)
```

hyperparameter.update *hyperparameter.update*

Description

Update hyperparameters Update shape (alpha) and scale (beta) parameters of the inverse gamma distribution.

Usage

```
hyperparameter.update(dat, alpha, beta, th = 0.01)
```

Arguments

dat	A probes x samples matrix (probeset).
alpha	Shape parameter of inverse gamma density for the probe variances.
beta	Scale parameter of inverse gamma density for the probe variances.
th	Convergence threshold.

Details

Shape update: $\alpha \leftarrow \alpha + T/2$; Scale update: $\beta \leftarrow \alpha * s2$ where $s2$ is the updated variance for each probe (the mode of variances is given by β/α). The variances ($s2$) are updated by EM type algorithm, see `s2.update`.

Value

A list with elements alpha, beta (corresponding to the shape and scale parameters of inverse gamma distribution, respectively).

Author(s)

Leo Lahti <leo.lahti@iki.fi>

References

See citation("RPA")

See Also

s2.update, rpa.online

Examples

```
#
## Generate and fit toydata, learn hyperparameters
#set.seed(11122)
#P <- 11 # number of probes
#N <- 5000 # number of arrays
#real <- sample.probeset(P = P, n = N, shape = 3, scale = 1, mu.real = 4)
#dat <- real$dat # probes x samples#
#
## Set priors
#alpha <- 1e-2
#beta <- rep(1e-2, P)
## Operate in batches
#step <- 1000
#for (ni in seq(1, N, step)) {
#  batch <- ni:(ni+step-1)
#  hp <- hyperparameter.update(dat[,batch], alpha, beta, th = 1e-2)
#  alpha <- hp$alpha
#  beta <- hp$beta
#}
## Final variance estimate
#s2 <- beta/alpha
#
## Compare real and estimated variances
#plot(sqrt(real$tau2), sqrt(s2), main = cor(sqrt(real$tau2), sqrt(s2))); abline(0,1)
```

levelmap

Map taxonomic levels

Description

Map taxa between hierarchy levels.

Usage

```
levelmap(taxa = NULL, from, to, tax.table)
```

Arguments

taxa	taxa to convert; if NULL then considering all taxa in the tax.table
from	convert from taxonomic level
to	convert to taxonomic level
tax.table	tax.table

Value

mappings

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

See citation('microbiome')

n.phylotypes.per.oligo

n.phylotypes.per.oligo

Description

Check number of matching phylotypes for each probe

Usage

```
n.phylotypes.per.oligo(taxonomy, level)
```

Arguments

taxonomy	oligo - phylotype matching data.frame
level	phylotype level

Value

number of matching phylotypes for each probe

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

See citation("microbiome")

online.quantile	<i>online.quantile</i> Quantile normalization tools for online preprocessing. Estimate quantiles for quantile normalization based on subset of the data (random, or specified by the user).
-----------------	---

Description

online.quantile Quantile normalization tools for online preprocessing. Estimate quantiles for quantile normalization based on subset of the data (random, or specified by the user).

Usage

```
online.quantile(abatch, n)
```

Arguments

abatch	AffyBatch
n	Numeric: number of random samples to use to define quantile basis. Vector: specify samples to be used in quantile basis calculation.

Details

"online.quantile": Ordinary quantile normalization is exhaustively memory-consuming in large data sets. Then the quantiles can be calculated based on subset of the data to allow efficient normalization. This function can also be used to investigate effect of subset size to convergence of the quantile estimates;"qnorm.basis.online": sweeps through the data in batches to calculate the basis for quantile normalization (average over sorted profiles).

Value

"online.quantile": AffyBatch; "qnorm.basis.online": a vector containing the basis for quantile normalization.

Author(s)

Leo Lahti <leo.lahti@iki.fi>

References

See citation("RPA")

Examples

```
#
```

```
probe.parameters.tolist
```

probe.parameters.tolist

Description

Convert probe parameter table into a list format

Usage

```
probe.parameters.tolist(probe.parameters)
```

Arguments

```
probe.parameters
```

A data.frame with alpha, betas, tau2, affinities, quantile.basis

Author(s)

Leo Lahti <leo.lahti@iki.fi>

References

See citation("RPA")

Examples

```
# df <- probe.parameters.tolist(probe.parameters.table)
```

```
probe.performance
```

Probe performance

Description

Provide a table of probe-level parameter estimates (affinity and stochastic noise) for RPA output.

Usage

```
probe.performance(probe.parameters, abatch, sets = NULL)
```

Arguments

```
probe.parameters
```

List with affinities and variances for the probesets

```
abatch
```

Affybatch used in the analysis

```
sets
```

Specify the probesets to include in the output. Default: All probesets

Value

Data frame of probe-level parameter estimates

Author(s)

Leo Lahti <leo.lahti@iki.fi>

References

See citation("RPA")

probeplot	<i>probeplot</i> Plot RPA results and probe-level data for a specified probeset.
-----------	--

Description

probeplot Plot RPA results and probe-level data for a specified probeset.

Usage

```
probeplot(
  dat,
  highlight.probes = NULL,
  pcol = "darkgrey",
  hcol = "red",
  cex.lab = 1.5,
  cex.axis = 1,
  cex.main = 1,
  cex.names = 1,
  main = "",
  ...
)
```

Arguments

dat	Background-corrected and normalized data: probes x samples.
highlight.probes	Optionally highlight some of the probes (with dashed line)
pcol	Color for probe signal visualization.
hcol	Color for probe highlight
cex.lab	Label size adjustment parameters.
cex.axis	Axis size adjustment parameters.
cex.main	Title size adjustment parameters.
cex.names	Names size adjustment parameters.
main	Title text.
...	Other parameters to pass for plot function.

Details

Plots the preprocessed probe-level observations, estimated probeset-level signal, and probe-specific variances. It is also possible to highlight individual probes and external summary measures.

Value

Used for its side-effects. Returns probes x samples matrix of probe-level data plotted on the image.

Author(s)

Leo Lahti <leo.lahti@iki.fi>

References

See citation("RPA")

Examples

```
#
```

probetable

probetable

Description

Convert probe-level hyperparameter lists into a table format.

Usage

```
probetable(probe.parameters)
```

Arguments

probe.parameters

A list with alpha, betas, variances and affinities

Author(s)

Leo Lahti <leo.lahti@iki.fi>

References

See citation("RPA")

Examples

```
# df <- probetable(probe.parameters)
```

retrieve.probesets	<i>Retrieve probesets</i>
--------------------	---------------------------

Description

List probes for each probeset in taxonomic data.

Usage

```
retrieve.probesets(tax.table, level = "species", name = NULL)
```

Arguments

tax.table	data.frame with oligo - phylotype mapping info
level	phylotype level for probesets
name	specify phylotypes to check (optional)

Value

A list. Probes for each phylotype.

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

See citation('microbiome')

Examples

```
#tax.table <- GetPhylogeny('HITChip')
#sets <- retrieve.probesets(tax.table, 'species', 'Weissella confusa')
```

rpa	<i>rpa</i>
-----	------------

Description

Wrapper for RPA preprocessing.

Usage

```
rpa(
  abatch = NULL,
  verbose = FALSE,
  bg.method = "rma",
  normalization.method = "quantiles.robust",
  cdf = NULL,
  cel.files = NULL,
  cel.path = NULL,
  probe.parameters = NULL,
  mc.cores = 1,
  summarize.with.affinities = FALSE
)
```

Arguments

<code>abatch</code>	An AffyBatch object.
<code>verbose</code>	Print progress information during computation.
<code>bg.method</code>	Specify background correction method. Default: "rma". See <code>bgcorrect.methods()</code> for other options.
<code>normalization.method</code>	Specify quantile normalization method. Default: "pmonly". See <code>normalize.methods(Dilution)</code> for other options.
<code>cdf</code>	Specify an alternative CDF environment. Default: none.
<code>cel.files</code>	List of CEL files to preprocess.
<code>cel.path</code>	Path to CEL file directory.
<code>probe.parameters</code>	A list, each element corresponding to a probe set. Each probeset element has the following optional elements: mu (affinity), tau2 (variance), alpha (shape prior), beta (scale prior). Each of these elements contains a vector over the probeset probes, specifying the probe parameters according to the RPA model. If variance is given, it overrides the priors. Can be also used to set user-specified priors for the model parameters. Not used tau2.method = "var". The prior parameters alpha and beta are prior parameters for inverse Gamma distribution of probe-specific variances. Noninformative prior is obtained with alpha, beta -> 0. Not used with tau2.method 'var'. Scalar alpha and beta specify an identical inverse Gamma prior for all probes, which regularizes the solution. Can be also specified as lists, each element corresponding to one probeset. May also include <code>quantile.basis</code>
<code>mc.cores</code>	Number of cores for parallelized processing.
<code>summarize.with.affinities</code>	Use affinity estimates in probe summarization step. Default: FALSE.

Details

RPA preprocessing function. Gives an estimate of the probeset-level mean parameter d of the RPA model, and returns these in an `expressionSet` object. The choices `tau2.method = "robust"` and `d.method = "fast"` are recommended. With small sample size and informative prior, `d.method = "basic"` may be preferable. For very large expression data collections, see `rpa.online` function.

Value

Preprocessed expression matrix in expressionSet format

Author(s)

Leo Lahti <leo.lahti@iki.fi>

References

See citation("RPA")

See Also

rpa.online, AffyBatch, ExpressionSet, estimate.affinities, rpa.fit

Examples

```
# eset <- rpa(abatch)
```

rpa.complete	<i>Complete RPA preprocessing</i>
--------------	-----------------------------------

Description

RPA preprocessing, also returns probe parameters.

Usage

```
rpa.complete(  
  abatch = NULL,  
  sets = NULL,  
  epsilon = 0.01,  
  tau2.method = "robust",  
  d.method = "fast",  
  verbose = FALSE,  
  bg.method = "rma",  
  normalization.method = "quantiles.robust",  
  cdf = NULL,  
  cel.files = NULL,  
  cel.path = NULL,  
  probe.parameters = list(),  
  mc.cores = 1,  
  summarize.with.affinities = FALSE  
)
```

Arguments

<code>abatch</code>	An AffyBatch object.
<code>sets</code>	Probesets for which RPA will be computed.
<code>epsilon</code>	Convergence tolerance. The iteration is deemed converged when the change in all parameters is $< \epsilon$.
<code>tau2.method</code>	Optimization method for tau2 (probe-specific variances). This parameter is denoted by τ^2 in the vignette and manuscript "robust": (default) update tau2 by posterior mean, regularized by informative priors that are identical for all probes (user-specified by setting scalar values for alpha, beta). This regularizes the solution, and avoids overfitting where a single probe obtains infinite reliability. This is a potential problem in the other tau2 update methods with non-informative variance priors. The default values $\alpha = 2$; $\beta = 1$ are used if alpha and beta are not specified. "mode": update tau2 with posterior mean "mean": update tau2 with posterior mean "var": update tau2 with variance around d. Applies the fact that tau2 cost function converges to variance with large sample sizes.
<code>d.method</code>	Method to optimize d. "fast": (default) weighted mean over the probes, weighted by probe variances. The solution converges to this with large sample size. "basic": optimization scheme to find a mode used in Lahti et al. TCBB/IEEE; relatively slow; this is the preferred method with small sample sizes.
<code>verbose</code>	Print progress information during computation.
<code>bg.method</code>	Specify background correction method. Default: "rma". See <code>bgcorrect.methods()</code> for other options.
<code>normalization.method</code>	Specify quantile normalization method. Default: "pmonly". See <code>normalize.methods(Dilution)</code> for other options.
<code>cdf</code>	Specify an alternative CDF environment. Default: none.
<code>cel.files</code>	List of CEL files to preprocess.
<code>cel.path</code>	Path to CEL file directory.
<code>probe.parameters</code>	A list, each element corresponding to a probe set. Each probeset element has the following optional elements: affinity (affinity), tau2 (variance), alpha (shape prior), betas (scale prior). Each of these elements contains a vector over the probeset probes, specifying the probe parameters according to the RPA model. If variance is given, it overrides the priors. Can be also used to set user-specified priors for the model parameters. Not used <code>tau2.method = "var"</code>. The prior parameters alpha and beta are prior parameters for inverse Gamma distribution of probe-specific variances. Noninformative prior is obtained with $\alpha, \beta \rightarrow 0$. Not used with <code>tau2.method 'var'</code>. Scalar alpha and beta specify an identical inverse Gamma prior for all probes, which regularizes the solution. Can be also specified as lists, each element corresponding to one probeset. Can also include <code>quantile.basis</code>
<code>mc.cores</code>	Number of cores for parallelized processing.
<code>summarize.with.affinities</code>	Use affinity estimates in probe summarization step. Default: FALSE.

Details

RPA preprocessing function. Gives an estimate of the probeset-level mean parameter d of the RPA model, and returns these in an expressionSet object. The choices `tau2.method = "robust"` and `d.method = "fast"` are recommended. With small sample size and informative prior, `d.method = "basic"` may be preferable. For very large expression data collections, see `rpa.online` function.

Value

List with preprocessed expression matrix, corresponding probe parameters, AffyBatch and CDF

Author(s)

Leo Lahti <leo.lahti@iki.fi>

References

See `citation("RPA")`

Examples

```
# eset <- rpa(abatch)
```

rpa.fit	<i>RPA fit</i>
---------	----------------

Description

Fit the RPA model.

Usage

```
rpa.fit(
  dat,
  epsilon = 0.01,
  alpha = NULL,
  beta = NULL,
  tau2.method = "robust",
  d.method = "fast",
  summarize.with.affinities = FALSE
)
```

Arguments

dat	Original data: probes x samples.
epsilon	Convergence tolerance. The iteration is deemed converged when the change in all parameters is < epsilon.
alpha	alpha prior for inverse Gamma distribution of probe-specific variances. Non-informative prior is obtained with alpha, beta -> 0. Not used with tau2.method 'var'. Scalar alpha and beta are specify equal inverse Gamma prior for all probes to regularize the solution. The defaults depend on the method.

beta	beta prior for inverse Gamma distribution of probe-specific variances. Noninformative prior is obtained with alpha, beta -> 0. Not used with tau2.method 'var'. Scalar alpha and beta are specify equal inverse Gamma prior for all probes to regularize the solution. The defaults depend on the method.
tau2.method	Optimization method for tau2 (probe-specific variances); "robust": (default) update tau2 by posterior mean, regularized by informative priors that are identical for all probes (user-specified by setting scalar values for alpha, beta). This regularizes the solution, and avoids overfitting where a single probe obtains infinite reliability. This is a potential problem in the other tau2 update methods with non-informative variance priors. The default values alpha = 2; beta = 1 are used if alpha and beta are not specified. "mode": update tau2 with posterior mean "mean": update tau2 with posterior mean "var": update tau2 with variance around d. Applies the fact that tau2 cost function converges to variance with large sample sizes.
d.method	Method used to optimize d. Options: "fast": (default) weighted mean over the probes, weighted by probe variances. The solution converges to this with large sample size. "basic": optimization scheme to find a mode used in Lahti et al. TCBB/IEEE; relatively slow; preferred with small sample size.
summarize.with.affinities	Use affinity estimates in probe summarization step. Default: FALSE.

Details

Fits the RPA model, including estimation of probe-specific affinity parameters. First learns a point estimate for the RPA model in terms of differential expression values w.r.t. reference sample. After this, probe affinities are estimated by comparing original data and differential expression shape, and setting prior assumptions concerning probe affinities.

Value

mu: Fitted signal in original data: $\mu_{\text{real}} + d$; μ_{real} : Shifting parameter of the reference sample;
tau2: Probe-specific stochastic noise; affinity: Probe-specific affinities; data: Probeset data matrix;
alpha, beta: prior parameters

Author(s)

Leo Lahti <leo.lahti@iki.fi>

References

See citation("RPA")

See Also

rpa, estimate.affinities

Examples

```
# res <- rpa.fit(dat, epsilon, alpha, beta, tau2.method, d.method, affinity.method)
```

RPA.iteration	<i>RPA iteration</i>
---------------	----------------------

Description

Estimating model parameters d and τ_2 .

Usage

```
RPA.iteration(
  S,
  epsilon = 0.001,
  alpha = NULL,
  beta = NULL,
  tau2.method = "fast",
  d.method = "fast",
  maxloop = 1e+06
)
```

Arguments

<code>S</code>	Matrix of probe-level observations for a single probeset: samples x probes.
<code>epsilon</code>	Convergence tolerance. The iteration is deemed converged when the change in all parameters is $< \epsilon$.
<code>alpha</code>	alpha prior for inverse Gamma distribution of probe-specific variances. Non-informative prior is obtained with $\alpha, \beta \rightarrow 0$. Not used with <code>tau2.method 'var'</code> . Scalar alpha and beta are specify equal inverse Gamma prior for all probes to regularize the solution. The defaults depend on the method.
<code>beta</code>	beta prior for inverse Gamma distribution of probe-specific variances. Noninformative prior is obtained with $\alpha, \beta \rightarrow 0$. Not used with <code>tau2.method 'var'</code> . Scalar alpha and beta are specify equal inverse Gamma prior for all probes to regularize the solution. The defaults depend on the method.
<code>tau2.method</code>	Optimization method for τ_2 (probe-specific variances). "robust": (default) update τ_2 by posterior mean, regularized by informative priors that are identical for all probes (user-specified by setting scalar values for α, β). This regularizes the solution, and avoids overfitting where a single probe obtains infinite reliability. This is a potential problem in the other τ_2 update methods with non-informative variance priors. The default values $\alpha = 2$; $\beta = 1$ are used if α and β are not specified. "mode": update τ_2 with posterior mean "mean": update τ_2 with posterior mean "var": update τ_2 with variance around d . Applies the fact that τ_2 cost function converges to variance with large sample sizes.
<code>d.method</code>	Method to optimize d . "fast": (default) weighted mean over the probes, weighted by probe variances The solution converges to this with large sample size. "basic": optimization scheme to find a mode used in Lahti et al. TCBB/IEEE; relatively slow; this is the preferred method with small sample sizes.
<code>maxloop</code>	Maximum number of iterations in the estimation process.

Details

Finds point estimates of the model parameters d (estimated true signal underlying probe-level observations), and τ^2 (probe-specific variances). Assuming data set S with P observations of signal d with Gaussian noise that is specific for each observation (specified by a vector τ^2 of length P), this method gives a point estimate of d and τ^2 . Probe-level variance priors α , β can be used with τ^2 .methods 'robust', 'mode', and 'mean'. The d .method = "fast" is the recommended method for point computing point estimates with large samples size.

Value

A list with the following elements: d : A vector. Estimated 'true' signal underlying the noisy probe-level observations.; τ^2 : A vector. Estimated variances for each measurement (or probe).

Author(s)

Leo Lahti <leo.lahti@iki.fi>

References

See citation("RPA")

Examples

```
#
```

rpa.online

rpa.online

Description

RPA-online for preprocessing very large expression data sets.

Usage

```
rpa.online(
  cel.path = NULL,
  cel.files = NULL,
  sets = NULL,
  cdf = NULL,
  bg.method = "rma",
  probe.parameters = list(alpha = 1, beta = 1),
  epsilon = 0.01,
  mc.cores = 1,
  verbose = TRUE,
  shuffle = TRUE,
  batch.size = 100,
  batches = NULL,
  save.batches.dir = ".",
  keep.batch.files = FALSE,
  unique.run.identifier = paste("RPA-run-id-", rnorm(1), sep = ""),
  rseed = 23,
```

```

    speedup = TRUE,
    summarize.with.affinities = FALSE
  )

```

Arguments

<code>cel.path</code>	Path to CEL file directory
<code>cel.files</code>	List of CEL files to preprocess
<code>sets</code>	Probesets for which RPA will be computed
<code>cdf</code>	Specify an alternative CDF environment
<code>bg.method</code>	Specify background correction method. See <code>bgcorrect.methods()</code> for options.
<code>probe.parameters</code>	Can be used to set user-specified priors for the model parameters alpha, beta. Not used <code>tau2.method = "var"</code> . The prior parameters alpha and beta are prior parameters for inverse Gamma distribution of probe-specific variances. Noninformative prior is obtained with alpha, beta $\rightarrow 0$. Not used with <code>tau2.method 'var'</code> . Scalar alpha and beta specify an identical inverse Gamma prior for all probes, which regularizes the solution. Can be also specified as lists, each element corresponding to one probeset. May also include <code>quantile.basis</code> , which should be provided at log2 domain.
<code>epsilon</code>	Convergence tolerance. The iteration is deemed converged when the change in all parameters is $< \epsilon$.
<code>mc.cores</code>	Number of cores for parallel computation
<code>verbose</code>	Print progress information during computation
<code>shuffle</code>	Form random batches
<code>batch.size</code>	Batch size for online mode (<code>rpa.online</code>); the complete list of CEL files will be preprocessed in batches with this size using Bayesian online-updates for probe-specific parameters.
<code>batches</code>	User-defined CEL file batches
<code>save.batches.dir</code>	Output directory for temporary batch saves.
<code>keep.batch.files</code>	Logical. Keep (TRUE) or remove (FALSE) the batch files after preprocessing.
<code>unique.run.identifier</code>	Define identifier for this run for naming the temporary batch files. By default, a random id is generated.
<code>rseed</code>	Random seed.
<code>speedup</code>	Speed up computations with approximations.
<code>summarize.with.affinities</code>	Use affinity estimates in probe summarization step. Default: FALSE.

Details

`rpa.online` is used to preprocess very large expression data collections based on a Bayesian hyperparameter update procedure. Returns an `expressionSet` object preprocessed with RPA. Gives an estimate of the probeset-level mean parameter d of the RPA model, and returns these in an `expressionSet` object. The CEL files are handled in batches to obtain Bayesian updates for probe-specific hyperpriors; after sweeping through the database in batches the results are combined. The online mode is useful for preprocessing very large expression data sets where ordinary preprocessing algorithms fail, without compromises in modelling stage.

Value

List with two elements: an instance of the 'expressionSet' class and probe parameters. For probe.parameters contents, see the probe.parameters input argument.

Author(s)

Leo Lahti <leo.lahti@iki.fi>

References

See citation("RPA")

See Also

rpa, AffyBatch, ExpressionSet

Examples

```
# eset <- rpa.online(cel.file.path)
```

rpa.plot

rpa.plot

Description

Plot RPA results and probe-level data for a specified probeset.

Usage

```
rpa.plot(
  x,
  set,
  highlight.probes = NULL,
  pcol = "darkgrey",
  mucol = "black",
  ecol = "red",
  external.signal = NULL,
  main = NULL,
  plots = "all",
  ...
)
```

Arguments

x	Output from rpa.complete function
set	probeset
highlight.probes	mark probes for highlight
pcol	probe color
mucol	probeset signal color

<code>ecol</code>	external signal color
<code>external.signal</code>	external signal to be plotted on top
<code>main</code>	title
<code>plots</code>	plot type
<code>...</code>	other arguments to be passed

Details

Plots the preprocessed probe-level observations, estimated probeset-level signal, and probe-specific variances. It is also possible to highlight individual probes and external summary measures.

Value

Used for its side-effects. Returns probes x samples matrix of probe-level data plotted on the image.

Author(s)

Leo Lahti <leo.lahti@iki.fi>

References

See citation("RPA")

Examples

```
#
```

RPA.preprocess	<i>RPA preprocessing</i>
----------------	--------------------------

Description

Preprocess AffyBatch object for RPA.

Usage

```
RPA.preprocess(
  abatch,
  bg.method = "rma",
  normalization.method = "quantiles.robust",
  cdf = NULL,
  cel.files = NULL,
  cel.path = NULL,
  quantile.basis = NULL
)
```

Arguments

<code>abatch</code>	An AffyBatch object.
<code>bg.method</code>	Specify background correction method. See <code>bgcorrect.methods(abatch)</code> for options.
<code>normalization.method</code>	Specify normalization method. See <code>normalize.methods(abatch)</code> for options. For memory-efficient online version, use "quantiles.online".
<code>cdf</code>	The CDF environment used in the analysis.
<code>cel.files</code>	List of CEL files to preprocess.
<code>cel.path</code>	Path to CEL file directory.
<code>quantile.basis</code>	Optional. Basis for quantile normalization. NOTE: required in original, not log2 scale!

Details

Background correction, quantile normalization and log2-transformation for probe-level raw data in `abatch`. Then probe-level differential expression is computed between the specified 'reference' array (`cind`) and the other arrays. Probe-specific variance estimates are robust against the choice of reference array.

Value

`fcmat`: Probes x arrays preprocessed differential expression matrix. `cind`: Specifies which array in `abatch` was selected as a reference in calculating probe-level differential expression. `cdf`: The CDF environment used in the analysis. `set.inds`: Indices for probes in each probeset, corresponding to the rows of `fcmat`.

Author(s)

Leo Lahti <leo.lahti@iki.fi>

References

See `citation("RPA")`

Examples

```
#
```

<code>rpa.summarize</code>	<i>rpa.summarize</i>
----------------------------	----------------------

Description

RPA summarization.

Usage

```
rpa.summarize(dat, affinities, variances, summarize.with.affinities = FALSE)
```

Arguments

dat Original data: probes x samples.
 affinities Probe affinities
 variances Probe variances
 summarize.with.affinities Use affinity estimates in probe summarization step. Default: FALSE.

Details

Summarizes the probes in a probe set according to the RPA model based on the given affinity and variance parameters.

Value

A vector. Probeset-level summary signal.

Author(s)

Leo Lahti <leo.lahti@iki.fi>

References

See citation("RPA")

See Also

rpa

Examples

```
# res <- rpa.summarize(dat, affinities, variances, summarize.with.affinities = FALSE)
```

rpaplot

rpaplot Plot RPA results and probe-level data for a specified probeset.

Description

rpaplot Plot RPA results and probe-level data for a specified probeset.

Usage

```
rpaplot(
  dat,
  mu = NULL,
  tau2 = NULL,
  affinity = NULL,
  highlight.probes = NULL,
  pcol = "darkgrey",
  mucol = "black",
  ecol = "red",
  cex.lab = 1.5,
```

```

    cex.axis = 1,
    cex.main = 1,
    cex.names = 1,
    external.signal = NULL,
    main = "",
    plots = "all",
    ...
)

```

Arguments

<code>dat</code>	Background-corrected and normalized data: probes x samples.
<code>mu</code>	probeset signal
<code>tau2</code>	probe variances
<code>affinity</code>	probe affinities
<code>highlight.probes</code>	Optionally highlight some of the probes (with dashed line)
<code>pcol</code>	Color for probe signal visualization.
<code>mucol</code>	Color for summary estimate.
<code>ecol</code>	Color for external signal.
<code>cex.lab</code>	Label size adjustment parameters.
<code>cex.axis</code>	Axis size adjustment parameters.
<code>cex.main</code>	Title size adjustment parameters.
<code>cex.names</code>	Names size adjustment parameters.
<code>external.signal</code>	Plot external signal on the probeset. For instance, an alternative summary estimate from another preprocessing methods
<code>main</code>	Title text.
<code>plots</code>	"all": plot data and summary, noise and affinity; "data": plot data and summary
<code>...</code>	Other parameters to pass for plot function.

Details

Plots the preprocessed probe-level observations, estimated probeset-level signal, and probe-specific variances. It is also possible to highlight individual probes and external summary measures.

Value

Used for its side-effects. Returns probes x samples matrix of probe-level data plotted on the image.

Author(s)

Leo Lahti <leo.lahti@iki.fi>

References

See citation("RPA")

Examples

```
#
```

sample.probeset	<i>sample.probeset</i>
-----------------	------------------------

Description

Toydata generator for probeset data.

Usage

```
sample.probeset(P = 10, n = 20, shape = 1, scale = 1, mu.real = 2)
```

Arguments

P	Number of probes.
n	Number of samples.
shape	Shape parameter of the inverse Gamma function used to generate the probe-specific variances.
scale	Scale parameters of the inverse Gamma function used to generate the probe-specific variances.
mu.real	Absolute signal level of the probeset.

Details

Generate random probeset with varying probe-specific affinities and variances. The toy data generator follows distributional assumptions of the RPA model and allows quantitative estimation of model accuracy with different options, noise levels and sample sizes. Probeset-level summary estimate is obtained as $\mu.\text{real} + d$.

Value

A list with the following elements:

dat	Probeset data: probes x samples
tau2	Probe variances.
affinity	Probe affinities.
d	Probeset signal shape.
mu.real	Probeset signal level.
mu	Probeset-level total signal.

Author(s)

Leo Lahti <leo.lahti@iki.fi>

References

See citation("RPA")

Examples

```
# real <- sample.probeset(P = 10, n = 20, shape = 1, scale = 1, mu.real = 2)
```

summarize.batch

summarize.batch

Description

Summarize batch.

Usage

```
summarize.batch(
  q,
  set.inds,
  probe.parameters = list(),
  epsilon,
  verbose = FALSE,
  mc.cores = 1,
  summarize.with.affinities = FALSE
)
```

Arguments

<code>q</code>	Background corrected, quantile-normalized, log2 probes x samples matrix
<code>set.inds</code>	Indices for each probeset, corresponding to <code>q</code> matrix
<code>probe.parameters</code>	A list, each element corresponding to a probe set. Each probeset element has the following elements: affinity, variance and optionally alpha and beta priors. Each of these elements contains a vector over the probeset probes, specifying the probe parameters according to the RPA model. If variances are given, that overrides the priors.
<code>epsilon</code>	Convergence tolerance. The iteration is deemed converged when the change in all parameters is < epsilon.
<code>verbose</code>	Print progress information during computation.
<code>mc.cores</code>	Number of cores for parallel processing
<code>summarize.with.affinities</code>	Use affinity estimates in probe summarization step. Default: FALSE.

Author(s)

Leo Lahti <leo.lahti@iki.fi>

References

See citation("RPA")

Examples

```
#
```

summarize.batches *summarize.batches*

Description

Summarize batches.

Usage

```
summarize.batches(
  sets = NULL,
  probe.parameters = list(),
  batches,
  load.batches = FALSE,
  mc.cores = 1,
  cdf = NULL,
  bg.method = "rma",
  normalization.method = "quantiles",
  verbose = TRUE,
  save.batches.dir = ".",
  unique.run.identifier = NULL,
  save.batches = FALSE,
  set.inds,
  speedup = FALSE,
  summarize.with.affinities = FALSE
)
```

Arguments

sets	Probesets to summarize
probe.parameters	Optional probe parameters, including priors.
batches	Data batches for online learning
load.batches	Logical. Load precalculated data for the batches.
mc.cores	Number of cores for parallel computation
cdf	CDF for alternative probeset definitions
bg.method	Background correction method
normalization.method	Normalization method
verbose	Print progress information
save.batches.dir	Specify the output directory for temporary batch saves.
unique.run.identifier	Define identifier for this run for naming the temporary batch files. By default, a random id is generated.
save.batches	Save batches?
set.inds	Probeset indices
speedup	Speed up calculations with approximations.
summarize.with.affinities	Use affinity estimates in probe summarization step. Default: FALSE.

Details

Sweeps through the batches. Summarizes the probesets within each batch based on the precalculated model parameter point estimates.

Value

Expression matrix: probesets x samples.

Author(s)

Leo Lahti <leo.lahti@iki.fi>

References

See citation("RPA")

Examples

```
#
```

summarize.rpa	<i>RPA summarization</i>
---------------	--------------------------

Description

Probeset summarization with RPA for taxonomic data.

Usage

```
summarize.rpa(
  taxonomy,
  level,
  probedata,
  verbose = TRUE,
  probe.parameters = NULL
)
```

Arguments

taxonomy	oligo - phylotype matching data.frame
level	taxonomic level for the summarization.
probedata	preprocessed probes x samples data matrix in absolute domain
verbose	print intermediate messages
probe.parameters	Optional. If probe.parameters are given, the summarization is based on these and model parameters are not estimated. A list. One element for each probeset with the following probe vectors: affinities, variances

Value

List with two elements: abundance.table (summarized data matrix in absolute scale) and probe.parameters (RPA probe level parameter estimates)

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

See citation("microbiome")

`summarize.sum`*Sum-based probe summarization*

Description

Probeset summarization with the standard sum method.

Usage

```
summarize.sum(  
  taxonomy,  
  level,  
  probedata,  
  verbose = TRUE,  
  downweight.ambiguous.probes = TRUE  
)
```

Arguments

<code>taxonomy</code>	oligo - phylotype matching data.frame
<code>level</code>	taxonomic level for the summarization.
<code>probedata</code>	preprocessed probes x samples data matrix in absolute domain
<code>verbose</code>	print intermediate messages
<code>downweight.ambiguous.probes</code>	Downweight probes with multiple targets

Value

List with two elements: `abundance.table` (summarized data matrix in absolute scale) and `probe.parameters` used in the calculations

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

See citation("microbiome")

summarize_probedata	<i>Summarize probedata</i>
---------------------	----------------------------

Description

Summarize phylogenetic microarray probe-level data from given input folder.

Usage

```
summarize_probedata(
  data.dir = NULL,
  probedata = NULL,
  taxonomy = NULL,
  level,
  method,
  probe.parameters = NULL
)
```

Arguments

data.dir	Data folder.
probedata	probe-level data matrix in absolute domain
taxonomy	probe taxonomy
level	Summarization level
method	Summarization method
probe.parameters	Precalculater probe parameters. Optional.

Value

data matrix (taxa x samples)

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

See citation('microbiome')

Examples

```
## Not run:
#library(microbiome)
#data.directory <- system.file("extdata", package = "microbiome")
# Read oligo-level data (here: simulated example data)
#probedata <- read_hitchip(data.directory, method = "frpa")$probedata
# Read phylogeny map
# NOTE: use phylogeny.filtered for species/L1/L2 summarization
# Load taxonomy from output directory
#taxonomy <- GetPhylogeny("HITChip", "filtered")
```

```
# Summarize oligos into higher level phylotypes
#dat <- summarize_probedata(
#      probedata = probedata,
#      taxonomy = taxonomy,
#      method = "rpa",
#      level = "species")
#
## End(Not run)
```

updating.hyperparameters

updating hyperparameters

Description

Hyperparameter update.

Usage

```
updating.hyperparameters(
  q,
  set.inds,
  verbose,
  mc.cores = 1,
  alpha,
  betas,
  epsilon
)
```

Arguments

q	probes x samples matrix
set.inds	Probe set indices
verbose	Print progress information
mc.cores	Number of cores for parallel computation
alpha	alpha hyperparameter
betas	beta hyperparameters
epsilon	Convergence parameter

Value

List with the following elements: alpha, betas, s2s (variances)

Author(s)

Leo Lahti <leo.lahti@iki.fi>

References

See citation("RPA")

Examples

#

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