# Package 'BUS'

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Author Yin Jin, Hesen Peng, Lei Wang, Raffaele Fronza, Yuanhua Liu and Christine Nardini
Maintainer Yuanhua Liu <li>liuyuanhua@picb.ac.cn&gt;</li>
<b>Description</b> This package can be used to compute associations among genes (genenetworks) or between genes and some external traits (i.e. clinical).
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BUS-package

For network reconstruction.

# **Description**

This package can be used to compute associations among genes (gene-networks) or between genes and some external traits (i.e. clinical). [Function: BUS]

Both associations can be computed via correlation or mutual information (MI). [Functions: gene.similarity (gene-gene associations) and gene.trait.similarity (gene-trait associations)]

Statistical significance of the association is computed for single and multiple hypotheses testing, using random permutations method [Functions: gene.pvalue, gene.trait.pvalue]

The package can handle data with missing values using bootstrapping methods to fill NAs. [Arguments: na.replica]

#### **Details**

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Type: Package
Version: 1.0.2
Date: 2009-10-31
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# Author(s)

Yin Jin, Hesen Peng, Lei Wang, Raffaele Fronza, Yuanhua Liu and Christine Nardini Maintainer: Yuanhua Liu</br>

BUS

A wrapper function for matrices of p-value and predicted network

# Description

A wrapper function to calculate the computation of two types of similarities (correlation and mutual information) with two different goals: (i) identification of the statistically significant similarities among the activity of molecules sampled across different experiments (option Unsupervised, U), (ii) identification of the statistically significant similarities between such molecules and other types of information (clinical etc., option supervised, S).

# Usage

BUS(EXP, trait = NULL, measure, method.permut = 2, n.replica = 400, net.trim = NULL, thresh = NULL, n

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## **Arguments**

EXP Gene expression data in form of a matrix. Row stands for genes and column for

experiments.

trait Trait data in form of a matrix. The row stands for traits and column for experi-

ments.

measure Metric used to calculate similarity: "corr" for correlation, "MI" for mutual in-

formation.

method.permut A flag to indicate which method is used to correct permutation p-values, default

as 2. See gene.pvalue for details.

n.replica Number of permutations used for the correction of multiple hypothesis testing;

default value is 400.

net.trim Method used to trim the network: "mrnet", "clr", "aracne" and "none". "mrnet"

infers a network using the maximum relevance/minimum redundancy feature selection method; "clr" use the CLR algorithm; "aracne" applies the data processing inequality to all triplets of nodes in order to remove the least significant edge in each triplet. These options come from the package *minet*, and they are used only for mutual information. "none" indicates no trim operation. It should

be chosen when correlation is considered.

thresh Threshold for significance of the corrected p-value. It is used, in the Unsuper-

vised case, to trim the adjacency matrix (contains the results of the gene-gene association based on the chosen metric) and obtain a predicted gene interaction network. In the Supervised case, since no network is predicted, it is set as

NULL.

nflag A flag to indicate a gene-gene interaction case (Unsupervised) or a gene-trait

interaction case (Supervised); 1 for Unsupervised and 2 for Supervised.

#### Value

similarity A matrix of similarity, which could be correlation or mutual information single.perm.p.value

A matrix of single p-values

multi.perm.p.value

A matrix of corrected p-values

net.pred.permut

Predicted network obtained trimming non-significant values

# Author(s)

Yin Jin, Hesen Peng, Lei Wang, Raffaele Fronza, Yuanhua Liu and Christine Nardini

### See Also

```
gene.pvalue,gene.trait.pvalue,pred.network
```

```
data(copasi)
mat<-as.matrix(copasi[1:10,])
rownames(mat)<-paste("G",1:nrow(mat), sep="")
BUS(EXP=mat,measure="corr",net.trim="none",thresh=0.05,nflag=1)</pre>
```

4 gene.pvalue

# **Description**

This dataset is taken from Copasi2 (Complex Pathway Simulator), a software for simulation and analysis of biochemical networks. The system generates random artificial gene networks according to well-defined topological and kinetic properties. These are used to run in silico experiments simulating real laboratory micro-array experiments. Noise with controlled properties is added to the simulation results several times emulating measurement replicates, before expression ratios are calculated. This series consists of 150 artificial gene networks. Each network consists of 100 genes with a total of 200 gene interactions (on average each gene has 2 modulators).

#### **Format**

A data frame is size of 100x100, the 100 rows represent 100 genes and 100 columns for 100 experiments.

#### References

See http://www.comp-sys-bio.org/AGN/data.html for detailed information.

8	gene.pvalue	Calculates p-value for gene-gene interaction

# **Description**

To calculate p-value for the null hypothesis that there is no gene-gene interaction. For gene expression data with M genes, a p-value matrix under MxM single null hypotheses (each two genes have no interaction) is computed; besides, matrices with correct p-values are output: corrected permutation method using a distribution of MxMxP (P number of permutations) null hypotheses tests (multi.perm.p.value). p-values are calculated based on the adjacency matrix for gene-gene interaction computed by function *gene.similarity*.

# Usage

```
gene.pvalue(EXP, measure, net.trim, n.replica = 400)
```

# **Arguments**

EXP	Gene expression data in form of a matrix. Row stands for genes and column for experiments.
measure	Metric used to calculate similarity between genes: "corr" for correlation, "MI" for mutual information.
net.trim	Method used to trim the network: "mrnet", "clr", "aracne" and "none". "mrnet" infers a network using the maximum relevance/minimum redundancy feature selection method; "clr" use the CLR algorithm; "aracne" applies the data processing inequality to all triplets of nodes in order to remove the least significant edge in each triplet. These options come from the package <i>minet</i> , and they are used only for mutual information. "none" indicates no trim operation. It should be chosen when correlation is considered.

gene.similarity 5

n.replica

Number of permutations used for the correction of multiple hypothesis testing; default value is 400.

#### **Details**

Normally, in a permutation method, we use the empirical distribution of some statistics to estimate the p-value. To get a simple p-value for no interaction between gene i and j, empirical distribution of a vector with length of P (number of replicates) is used; to correct for multiple hypothesis with permutations, an empirical distribution of a vector with length of PxM (M being the number of hypotheses tested) is used.

### Value

```
single.perm.p.value
```

A matrix of single p-values obtained with permutation method + beta distribution for extreme values (for MI) or obtained with the exact distribution computed directly by *cor.test* (for correlation)

multi.perm.p.value

A matrix of corrected p-values obtained with permutation method

### Author(s)

Yin Jin, Hesen Peng, Lei Wang, Raffaele Fronza, Yuanhua Liu and Christine Nardini

### See Also

```
gene.similarity
```

# **Examples**

```
data(copasi)
mat=as.matrix(copasi)[1:10,]
rownames(mat)<-paste("G",1:nrow(mat), sep="")
gene.pvalue(mat,measure="MI",net.trim="mrnet")</pre>
```

gene.similarity

Calculate adjacency matrix for gene-gene interaction

## **Description**

To calculate an adjacency matrix for gene-gene interaction (using correlation/mutual information metric). For gene expression data with M genes and N experiments, the adjacency matrix is in size of MxM. It is optional to get a trimmed adjacency matrix according to the argument *net.trim*, i.e. *mrnet*, *clr* and *aracne* (from the package *minet*).

# Usage

```
gene.similarity(EXP, measure, net.trim, na.replica = 50)
```

6 gene.trait.pvalue

# **Arguments**

EXP Gene expression data in form of a matrix. Row stands for genes and column for experiments. measure Metric used to calculate similarity between genes: "corr" for correlation, "MI" for mutual information. net.trim Method used to trim the adjacency matrix: "mrnet", "clr", "aracne" and "none". "mrnet" infers a network using the maximum relevance/minimum redundancy feature selection method; "clr" use the CLR algorithm; "aracne" applies the data processing inequality to all triplets of nodes in order to remove the least significant edge in each triplet. These options come from the package minet, and they are used only for mutual information. "none" indicates no trim operation. It should be chosen when correlation is considered. na.replica Times of replication for filling NANs in the impute method; default value is 50. The (smooth) bootstrapping approach is used to give an estimation to missing

#### Value

An adjacency matrix in size of MxM with rows and columns both standing for genes. Element in row i and column j indicates the similarity between gene i and gene j.

#### Author(s)

Yin Jin, Hesen Peng, Lei Wang, Raffaele Fronza, Yuanhua Liu and Christine Nardini

# **Examples**

```
data(copasi)
mat=as.matrix(copasi)[1:10,]
rownames(mat)<-paste("G",1:nrow(mat), sep="")
res<-gene.similarity(mat,measure="corr",net.trim="none")</pre>
```

value in the data.

gene.trait.pvalue

Calculate p-value for gene-trait interaction

# Description

To calculate p-value for null hypothesis that there is no interaction between gene and trait. There are MxT interactions between M genes and T traits. Results are given with 3 possibilities 1 for single p-value, and 3 for different types of correction. p-values are calculated based on the adjacency matrix for gene-gene interaction computed by function *gene.trait.similarity*.

# Usage

```
gene.trait.pvalue(EXP, trait, measure, method.permut = 2, n.replica = 400)
```

gene.trait.pvalue 7

## **Arguments**

EXP Gene expression data in form of a matrix. Row stands for genes and column for

experiments.

trait Trait data in form of matrix. Row stands for traits and column for experiments.

measure Metric used to calculate similarity: "corr" for correlation, "MI" for mutual in-

formation.

method.permut A flag to indicate correction style when multiple hypotheses testing is consid-

ered. 1 for multiple traits correction, 2 for multiple genes and 3 for both genes

and traits correction. The default value is 2.

n.replica Number of permutations for the correction of multiple hypothesis testing; de-

fault value is 400.

### **Details**

According to a permutation method, we use the empirical distribution of some statistics to estimate the p-value. For single p-value the empirical distribution is a vector of P (number of random replicates for each test) test values. It is then possible to correct p-value in different ways: method.permut = 1, it is the empirical distribution of a vector with length of TxP, corrects for the multiple traits tested; method.permut = 2, it is the empirical distribution of a vector with length of MxP, corrects for the multiple genes tested; method.permut = 3, it is empirical distribution of a vector with length of MxTxP, corrects for the multiple traits and genes tested.

#### Value

```
single.perm.p.value
```

A matrix of single p-values obtained with permutation method + beta distribution for extreme values (for MI) or obtained with the exact distribution computed directly by *cor.test* (for correlation)

multi.perm.p.value

A matrix of corrected p-values obtained with permutation method

# Author(s)

Yin Jin, Hesen Peng, Lei Wang, Raffaele Fronza, Yuanhua Liu and Christine Nardini

# See Also

```
gene.trait.similarity
```

```
data(tumors.mRNA)
data(tumors.miRNA)
exp<-tumors.mRNA
trait<-tumors.miRNA
gene.trait.pvalue(EXP=exp[1:10,],trait=trait[1:5,],measure="MI")</pre>
```

8 pred.network

```
gene.trait.similarity Calculate similarity for gene-trait interaction
```

# **Description**

To calculate similarity for gene-trait interaction (using correlation/mutual information metric).

### Usage

```
gene.trait.similarity(EXP, trait, measure, na.replica = 50)
```

in the data.

# **Arguments**

EXP	Gene expression data in form of a matrix. Row stands for genes and column for experiments.
trait	Trait data in form of matrix. Row stands for traits and column for experiments.
measure	Metric used to calculate similarity: "corr" for correlation, "MI" for mutual information.
na.replica	Times of replicates for filling NANs in impute method; default value is 50. The (smooth) bootstrapping approach is used to give an estimation to missing value

# Value

A matrix, row stands for gene and column for trait. Element in row i and column j stands for the association between the gene i and trait j.

# Author(s)

Yin Jin, Hesen Peng, Lei Wang, Raffaele Fronza, Yuanhua Liu and Christine Nardini

# **Examples**

```
data(tumors.mRNA)
data(tumors.miRNA)
exp<-tumors.mRNA
trait<-tumors.miRNA
gene.trait.similarity(EXP= exp[1:10, ],trait= trait[1:5, ],measure="MI")</pre>
```

pred.network

Predict the network

# Description

To predict the matrix of gene network, based on the similarity matrix and filtered according to a corrected p-value matrix.

## Usage

```
pred.network(pM,similarity,thresh)
```

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## **Arguments**

pM A corrected p-value matrix, a MxM matrix for significance of similarity among

M genes.

similarity A MxM matrix for similarity between genes. thresh Threshold for significance of the p-value.

#### Value

A MxM matrix of the predicted network, where cell emphij infers a link between gene i and j and set 0 when the p-value is not significant (no link).

#### Author(s)

Yin Jin, Hesen Peng, Lei Wang, Raffaele Fronza, Yuanhua Liu and Christine Nardini

# **Examples**

```
data(copasi)
mat<-as.matrix(copasi[1:10,])
rownames(mat)<-paste("G",1:nrow(mat), sep="")
similarity=gene.similarity(mat,measure="MI",net.trim="mrnet")
pM=gene.pvalue(mat,measure="MI",net.trim="mrnet")$single.perm.p.value
pred.network(pM,similarity,thresh=0.05)</pre>
```

tumors.miRNA

miRNA data from Human brain tumors

# **Description**

MiRNA data obtained by RT-PCR from human brain tumors. 12 brain tumors at different levels are analyzed for both mRNA and miRNA levels to study the correlation of any mRNA-miRNA pair in the *reference*.

# Usage

```
data(tumors.miRNA)
```

# **Format**

tumors.miRNA is a matrix with miRNA as rows and tumor type as columns.

#### References

Liu T, Papagiannakopoulos T, Puskar K, Qi S, Santiago F, Clay W, Lao K, Lee Y, Nelson SF, Kornblum HI, Doyle F, Petzold L, Shraiman B, Kosik KS. Detection of a microRNA signal in an in vivo expression set of mRNAs. Plos One. 2007; 2(8):e804.

```
data(tumors.miRNA)
tumors.miRNA[1:10,]
```

10 tumors.mRNA

tumors.mRNA

Gene expression data from Human brain tumors

# **Description**

Gene expression data obtained by microarray from human brain tumors. 12 brain tumors at different levels are analyzed for both mRNA and miRNA levels to study the correlation of any mRNA-miRNA pair in the *reference*.

# Usage

```
data(tumors.mRNA)
```

### **Format**

tumors.mRNA is a matrix with mRNA probe IDs as rows and tumor type as columns.

# References

Liu T, Papagiannakopoulos T, Puskar K, Qi S, Santiago F, Clay W, Lao K, Lee Y, Nelson SF, Kornblum HI, Doyle F, Petzold L, Shraiman B, Kosik KS. Detection of a microRNA signal in an in vivo expression set of mRNAs. Plos One. 2007; 2(8):e804.

```
data(tumors.mRNA)
tumors.mRNA[1:10,]
```

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