# Package 'intansv'

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Title	Integrative	analysis	of structural	l variations
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**Description** This package provides efficient tools to read and integrate structural variations predicted by popular softwares. Annotation and visulation of structural variations are also implemented in the package.

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**Depends** R (>= 2.14.0), plyr, ggbio, GenomicRanges

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methodsMerge	Integrate structural variations predicted by different methods

#### **Description**

Integrate predictions of different tools to provide more reliable structural variations.

## Usage

```
methodsMerge(..., others=NULL,
overLapPerDel = 0.8, overLapPerDup = 0.8, overLapPerInv = 0.8,
numMethodsSupDel = 2, numMethodsSupDup = 2, numMethodsSupInv = 2)
```

## **Arguments**

... results of different SVs predictions read in to R by intansv. others a data frame of structural variations predicted by other tools.

overLapPerDel Deletions predicted by different methods that have reciprocal coordinate overlap

larger than this threshold would be clustered together

overLapPerDup Duplications predicted by different methods that have reciprocal coordinate over-

lap larger than this threshold would be clustered together

overLapPerInv Inversions predicted by different methods that have reciprocal coordinate over-

lap larger than this threshold would be clustered together

numMethodsSupDel

Deletion clusters supportted by no more than this threshold of read support

would be discarded

numMethodsSupDup

Duplication clusters supportted by no more than this threshold of read support

would be discarded

numMethodsSupInv

Inversion clusters supportted by no more than this threshold of read support

would be discarded

#### **Details**

A structural variation (deletion, duplication, inversion et al.) may be reported by different tools. However, the boundaries of this structural variation predicted by different tools don't always agree with each other. Predictions of different methods with reciprocal overlap more than 80 percent were merged. Structural varions supported by only one method were discarded.

#### Value

A list with the following components:

del the integrated deletions of different methods.

dup the integrated duplications of different methods.

inv the integrated inversions of different methods.

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#### Author(s)

Wen Yao

#### **Examples**

```
breakdancer <- readBreakDancer(system.file("extdata/ZS97.breakdancer.sv",</pre>
                                 package="intansv"))
str(breakdancer)
cnvnator <- readCnvnator(system.file("extdata/cnvnator",package="intansv"))</pre>
str(cnvnator)
svseq <- readSvseq(system.file("extdata/svseq2",package="intansv"))</pre>
str(svseq)
delly <- readDelly(system.file("extdata/ZS97.DELLY.vcf",package="intansv"))</pre>
str(delly)
pindel <- readPindel(system.file("extdata/pindel",package="intansv"))</pre>
str(pindel)
sv_all_methods <- methodsMerge(breakdancer,pindel,cnvnator,delly,svseq)</pre>
str(sv_all_methods)
sv_all_methods.1 <- methodsMerge(breakdancer,pindel,cnvnator,delly,svseq,</pre>
                               overLapPerDel=0.7)
str(sv_all_methods.1)
sv_all_methods.2 <- methodsMerge(breakdancer,pindel,cnvnator,delly,svseq,</pre>
                               overLapPerDel=0.8, numMethodsSupDel=3)
str(sv_all_methods.2)
```

plotChromosome

Display the chromosome distribution of structural variations

#### **Description**

Display the chromosome distribution of structural variations by splitting the chromosomes into windows of specific size and counting the number of structural variations in each window.

# Usage

```
plotChromosome(genome, structuralVariation, windowSize=1000000)
```

#### **Arguments**

genome A data frame with ID and length of all Chromosomes.

structuralVariation

A list of structural variations.

windowSize A specific size (in base pair) to split chromosomes into windows.

plotRegion plotRegion

#### **Details**

To visualize the distribution of structural variations in the whole genome, chromosomes were splitted into windows of specific size (default 1 Mb) and the number of structural variations in each window were counted. The number of structural variations were shown using circular barplot.

#### Value

A circular plot with five layers:

- the circular view of genome ideogram.
- the chromosome coordinates labels.
- the circular barplot of number of deletions in each chromosome window.
- the circular barplot of number of duplications in each chromosome window.
- the circular barplot of number of inversions in each chromosome window.

#### Author(s)

Wen Yao

#### **Examples**

```
delly <- readDelly(system.file("extdata/ZS97.DELLY.vcf",package="intansv"))
str(delly)
genome.file.path <- system.file("extdata/chr05_chr10.genome.txt", package="intansv")
genome <- read.table(genome.file.path, head=TRUE, as.is=TRUE)
str(genome)
plotChromosome(genome,delly,1000000)</pre>
```

plotRegion

Display structural variations in a specific genomic region

## **Description**

Display the structural variations in a specific genomic region in circular view.

# Usage

plotRegion 5

#### **Arguments**

```
structuralVariation
```

A list of structural variations.

genomeAnnotation

A data frame of genome annotations.

regionChromosome

The chromosome identifier of a specific region to view.

regionStart The start coordinate of a specific region to view.

regionEnd The end coordinate of a specific region to view.

#### **Details**

Different SVs were shown as rectangles in different layers. See the package vignette and the example dataset for more details.

#### Value

A circular plot of all the structural variations and genes in a specific region with four layers:

- The composition of genes of a specific genomic region.
- The composition of deletions of a specific genomic region.
- The composition of duplications of a specific genomic region.
- The composition of inversions of a specific genomic region.

### Author(s)

Wen Yao

#### **Examples**

```
delly <- readDelly(system.file("extdata/ZS97.DELLY.vcf",package="intansv"))
str(delly)
anno.file.path <- system.file("extdata/chr05_chr10.anno.txt", package="intansv")
msu_gff_v7 <- read.table(anno.file.path, head=TRUE, as.is=TRUE)
str(msu_gff_v7)
plotRegion(delly,msu_gff_v7,"chr05",1,200000)</pre>
```

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readBreakDancer Read in the structural variations predicted by breakDancer
--

#### **Description**

Reading in the structural variations predicted by breakDancer, filtering low quality predictions and merging overlapping predictions.

## Usage

### **Arguments**

file the output file of breakDancer.

scoreCutoff the minimum score for a structural variation to be read in.

readsSupport the minimum read pair support for a structural variation to be read in.

regSizeLowerCutoff

the minimum size for a structural variation to be read in.

regSizeUpperCutoff

the maximum size for a structural variation to be read in.

method a tag to assign to the result of this function.

. . . parameters passed to read.table.

#### **Details**

The predicted SVs could be further filtered by score, number of read pairs supporting the occurence of a specific SV, and the predicted size of SVs to get more reliable SVs. See our paper for more details.

#### Value

A list with the following components:

del the deletions predicted by breakDancer.
inv the inversions predicted by breakDancer.

#### Author(s)

readCnvnator 7

### **Examples**

readCnvnator

Read in the structural variations predicted by CNVnator

## **Description**

Reading the structural variations predicted by CNVnator, filtering low quality predictions and merging overlapping predictions.

## Usage

### **Arguments**

dataDir the directory that contain the output files of CNVnator.

regSizeLowerCutoff

the minimum size for a structural variation to be read.

regSizeUpperCutoff

the maximum size for a structural variation to be read.

method a tag to assign to the result of this function.

#### **Details**

The predicted SVs could be further filtered by the predicted size of SVs to get more reliable SVs. See our paper for more details. The directory that specified by the parameter "dataDir" should only contain the predictions of CNVnator. See the example dataset for more details.

#### Value

A list with the following components:

del the deletions predicted by CNVnator.

dup the duplications predicted by CNVnator.

#### Author(s)

8 readDelly

#### **Examples**

```
cnvnator <- readCnvnator(system.file("extdata/cnvnator",package="intansv"))
str(cnvnator)</pre>
```

readDelly

Read in the structural variations predicted by DELLY

#### **Description**

Reading the structural variations predicted by DELLY, filtering low quality predictions and merging overlapping predictions.

#### Usage

```
readDelly(file="", regSizeLowerCutoff=100, regSizeUpperCutoff=1000000,
readsSupport=3, method="Delly", ...)
```

# **Arguments**

file the file containing the prediction results of DELLY.

regSizeLowerCutoff

the minimum size for a structural variation to be read.

regSizeUpperCutoff

the maximum size for a structural variation to be read.

readsSupport the minimum read pair support for a structural variation to be read.

method a tag to assign to the result of this function.

. . . parameters passed to read.table.

## **Details**

The predicted SVs could be further filtered by the number of read pairs supporting the occurence of a specific SV, and the predicted size of SVs to get more reliable SVs. See our paper for more details.

## Value

A list with the following components:

del the deletions predicted by DELLY.
dup the duplications predicted by DELLY.
inv the inversions predicted by DELLY.

## Author(s)

readLumpy 9

### **Examples**

```
delly <- readDelly(system.file("extdata/ZS97.DELLY.vcf",package="intansv"))
str(delly)</pre>
```

readLumpy

Read in the structural variations predicted by Lumpy

## Description

Reading the structural variations predicted by Lumpy, filtering low quality predictions and merging overlapping predictions.

#### Usage

```
\label{lem:continuous} readLumpy(file="", regSizeLowerCutoff=100, regSizeUpperCutoff=1000000, readsSupport=3, method="Lumpy", \ldots)
```

### **Arguments**

file the file containing the prediction results of Lumpy.

regSizeLowerCutoff

the minimum size for a structural variation to be read.

 ${\tt regSizeUpperCutoff}$ 

the maximum size for a structural variation to be read.

readsSupport the minimum read pair support for a structural variation to be read.

method a tag to assign to the result of this function.

... parameters passed to read.table.

#### **Details**

The predicted SVs could be further filtered by the number of reads supporting the occurrence of a specific SV, and the predicted size of SVs to get more reliable SVs. See our paper for more details.

#### Value

A list with the following components:

del the deletions predicted by Lumpy.
dup the duplications predicted by Lumpy.
inv the inversions predicted by Lumpy.

#### Author(s)

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### **Examples**

```
lumpy <- readLumpy(system.file("extdata/ZS97.LUMPY.vcf",package="intansv"))
str(lumpy)</pre>
```

readPindel

Read in the structural variations predicted by Pindel

#### **Description**

Reading the structural variations predicted by Pindel, filtering low quality predictions and merging overlapping predictions.

## Usage

## **Arguments**

dataDir the directory containing the prediction results of Pindel.

regSizeLowerCutoff

the minimum size for a structural variation to be read.

regSizeUpperCutoff

the maximum size for a structural variation to be read.

readsSupport the minimum read pair support for a structural variation to be read.

method a tag to assign to the result of this function.

#### Details

The predicted SVs could be further filtered by the number of reads supporting the occurence of a specific SV, and the predicted size of SVs to get more reliable SVs. See our paper for more details. The directory that specified by the parameter "dataDir" should only contain the predictions of Pindel. The deletions output files should be named using the suffix "\_D", the duplications output files should be named using the suffix "\_TD", and the inversions output files should be named using the suffix "\_INV". See the example dataset for more details.

#### Value

A list with the following components:

del the deletions predicted by Pindel.
dup the duplications predicted by Pindel.
inv the inversions predicted by Pindel.

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#### Author(s)

Wen Yao

#### **Examples**

```
pindel <- readPindel(system.file("extdata/pindel",package="intansv"))
str(pindel)</pre>
```

readSoftSearch

Read in the structural variations predicted by SoftSearch

# **Description**

Reading the structural variations predicted by SoftSearch, filtering low quality predictions and merging overlapping predictions.

#### Usage

#### **Arguments**

file the file containing the prediction results of SoftSearch.

regSizeLowerCutoff

the minimum size for a structural variation to be read.

regSizeUpperCutoff

the maximum size for a structural variation to be read.

readsSupport the minimum read pair support for a structural variation to be read.

method a tag to assign to the result of this function.

softClipsSupport

the minimum soft clip support for a structural variation to be read.

... parameters passed to read.table

### **Details**

The predicted SVs could be further filtered by the number of reads supporting the occurrence of a specific SV, and the predicted size of SVs to get more reliable SVs. See our paper for more details.

#### Value

A list with the following components:

del the deletions predicted by SoftSearch.
dup the duplications predicted by SoftSearch.
inv the inversions predicted by SoftSearch.

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#### Author(s)

Wen Yao

# **Examples**

```
softSearch <- readSoftSearch(system.file("extdata/ZS97.softsearch",package="intansv"))
str(softSearch)</pre>
```

readSvseq

Read in the structural variations predicted by SVseq2

#### **Description**

Reading the structural variations predicted by SVseq2, filtering low quality predictions and merging overlapping predictions.

## Usage

# Arguments

dataDir a directory containing the predictions of SVseq2.

 ${\tt regSizeLowerCutoff}$ 

the minimum size for a structural variation to be read.

 ${\tt regSizeUpperCutoff}$ 

the maximum size for a structural variation to be read.

readsSupport the minimum read pair support for a structural variation to be read.

method a tag to assign to the result of this function.

#### **Details**

The predicted SVs could be further filtered by the number of reads supporting the occurence of a specific SV, and the predicted size of SVs to get more reliable SVs. See our paper for more details. The directory that specified by the parameter "dataDir" should only contain the predictions of SVseq2. The deletions output files should be named using the suffix ".del". See the example dataset for more details.

#### Value

A list with the following components:

del the deletions predicted by SVseq2.

svAnnotation 13

#### Author(s)

Wen Yao

#### **Examples**

```
svseq <- readSvseq(system.file("extdata/svseq2",package="intansv"))
str(svseq)</pre>
```

svAnnotation

Annotation of structural variations

# Description

Annotate the effect caused by structural variations to genes and elements of genes.

## Usage

```
svAnnotation(structuralVariation,genomeAnnotation)
```

## **Arguments**

```
\label{eq:approx} A \ data \ frame \ of \ structural \ variations. genome Annotation
```

A data frame of genome annotations.

## **Details**

A structural variation (deletion, duplication, inversion et al.) could affect the structure of a specific gene, including deletion of introns/exons, deletion of whole gene, et al.. This function gives the detailed effects caused by structural variations to genes and elements of genes.

The parameter "structural Variation" should be a data frame with three columns:

- chromosome the chromosome of a structural variation.
- pos1 the start coordinate of a structural variation.
- pos2 the end coordinate of a structural variation.

#### Value

A data frame with the following columns:

chromosome	the chromosome of a structural variation.
pos1	the start coordinate of a structural variation.
pos2	the end coordinate of a structural variation.
size	the size of a structural variation.

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info information on a structural variation.

tag the tag of a genomic element.

start the start coordinate of a genomic element. end the end coordinate of a genomic element.

strand the strand of a genomic element.

ID the ID of a genomic element.

## Author(s)

Wen Yao

## **Examples**

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